Novel Inhibitors of Human Histone Deacetylases: Design, Synthesis, Enzyme Inhibition, and Cancer Cell Growth Inhibition of SAHA-Based Non-hydroxamates

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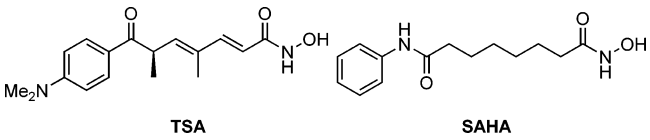
To find novel non-hydroxamate histone deacetylase (HDAC) inhibitors, a series of compounds modeled after suberoylanilide hydroxamic acid (SAHA) was designed and synthesized. In this series, compound **7**, in which the hydroxamic acid of SAHA is replaced by a thiol, was found to be as potent as SAHA, and optimization of this series led to the identification of HDAC inhibitors more potent than SAHA. In cancer cell growth inhibition assay, *S*-isobutyryl derivative **51** showed strong activity, and its potency was comparable to that of SAHA. The cancer cell growth inhibitory activity was verified to be the result of histone hyperacetylation and subsequent induction of $p21^{WAF1/CIP1}$ by Western blot analysis. Kinetical enzyme assay and molecular modeling suggest the thiol formed by enzymatic hydrolysis within the cell interacts with the zinc ion in the active site of HDACs.

Introduction

The reversible acetylation of the ϵ-amino groups of specific histone lysine residues by histone deacetylases (HDACs) and histone acetyl transferases is an important regulatory mechanism of gene expression.[1] When HDACs are inhibited, histone hyperacetylation occurs. The disruption of the chromatin structure by histone hyperacetylation leads to the transcriptional activation of a number of genes.[2] One important outcome of the activation is induction of the cyclin-dependent kinase inhibitory protein $p21^{WAF1/CIP1}$, which causes cell cycle arrest.[3] Indeed, HDAC inhibitors such as trichostatin A (TSA) and suberoylanilide hydroxamic acid (SAHA) (Chart 1) have been reported to inhibit cell growth, induce terminal differentiation in tumor cells,[4] and prevent the formation of malignant tumors in mice.[5] Therefore, HDACs have emerged as attractive targets in anticancer drug development, and HDAC inhibitors have also been viewed as useful tools to study the function of these enzymes.

Many groups have ongoing research programs to find nonpeptide small-molecule inhibitors of HDACs, and these efforts have led to the identification of several classes of inhibitors.[6] Most previously reported HDAC inhibitors belong to hydroxamic acid derivatives, typified by TSA and SAHA, which are thought to chelate the zinc ion in the active site in a bidentate fashion through its CO and OH groups.[7] However, hydroxamic acids occasionally have been associated with problems such as poor pharmacokinetics and severe toxicity.[8] Thus, it has become increasingly desirable to find

Chart 1



replacements that possess strong inhibitory action against HDACs. In addition, in terms of biological research, the discovery of novel zinc-binding groups (ZBGs) may lead to a new type of HDAC isozyme-selective inhibitors which are useful as tools for probing the biology of the enzyme.[9] Thus far, *o*-aminoanilide,[9,10] electrophilic ketones,[11] and *N*-formyl hydroxylamine[12] have been reported as ZBGs in small-molecule HDAC inhibitors. However, most of them have reduced potency as compared to hydroxamic acid, and unfortunately, HDAC inhibitors bearing electrophilic ketones[11] have a metabolic disadvantage in that they are readily reduced to inactive alcohols in vivo, even within cells. We therefore initiated a search for replacement groups for hydroxamic acid with the goal of drug discovery as well as finding new tools for biological research, and found some potent non-hydroxamate small-molecule HDAC inhibitors.[13] We now present a full account of our study reporting the design, synthesis, HDAC inhibition, cancer cell growth inhibition, and binding mode analysis of non-hydroxamates based on the structure of SAHA.

Chemistry

The compounds prepared for this study are shown in Tables 1−5. The routes used for synthesis of the compounds are shown in Schemes 1−4. Scheme 1 shows the preparation of compounds **4**, **5**, **10**, **12**−**17**, and **18**. Compounds **4** and **5** were synthesized from pimelic acid **56**. The condensation of pimelic acid **56** with an equiva-

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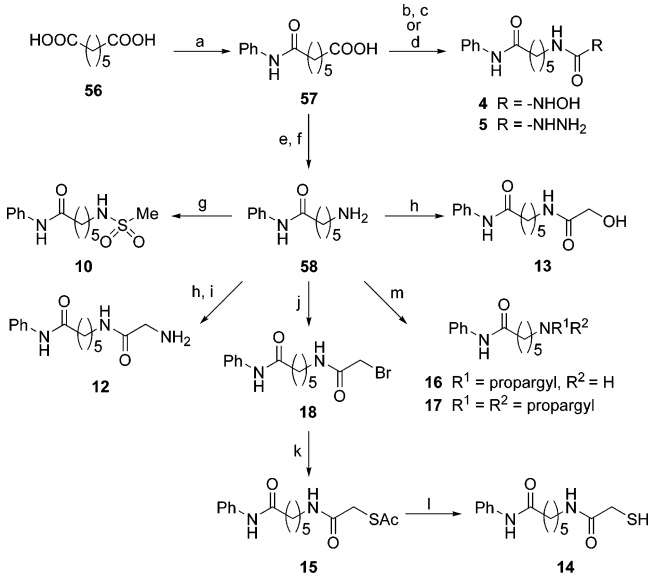
Scheme 1[a]



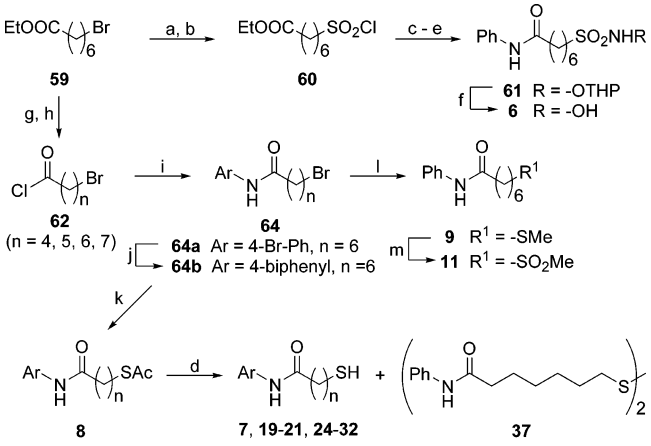
[a] Reagents and conditions: (a) aniline, 180 °C; (b) diphenylphosphoryl azide (DPPA), Et_3N, toluene, reflux, and then *O*-(2-tetrahydropyranyl)hydroxylamine, reflux; (c) TsOH, MeOH, rt; (d) DPPA, Et_3N, benzene, reflux, and then hydrazine monohydrate, reflux; (e) DPPA, Et_3N, benzene, reflux, and then BnOH, reflux; (f) H_2, 5%Pd−C, MeOH, rt; (g) MsCl, pyridine, rt; (h) $HOCH_2COOH$ or $BocNHCH_2COOH$, EDCI, HOBt, DMF, rt; (i) TFA, $CHCl_3$, rt; (j) $BrCH_2COBr$, Et_3N, THF, rt; (k) AcSK, EtOH, rt; (l) K_2CO_3, MeOH, rt; (m) propargyl bromide, K_2CO_3, MeOH, rt.

lent amount of aniline gave mono-anilide **57**. Curtius rearrangement of the acyl azide prepared from carboxylic acid **57** using diphenylphosphoryl azide provided the isocanates, which on treatment with *O*-tetrahydropyranyl (THP) hydroxylamine or hydrazine gave *O*-THP hydroxyurea and semicarbazide **5**. Deprotection of the THP group of the *O*-THP hydroxyurea under acidic conditions afforded hydroxyurea **4**.

Compounds **10**, **12**−**17**, and **18** were prepared from carboxylic acid **57** obtained above via amine **58** by the procedure outlined in Scheme 1. Carboxylic acid **57** was converted to amine **58** with a three-step sequence: Curtius rearrangement of the acyl azide prepared from carboxylic acid **57**, treatment of the resulting isocyanates with benzyl alcohol, and removal of the Z group by hydrogenation. Coupling between amine **58** and methanesulfonyl chloride afforded sulfonamide **10**. The reaction of amine **58** with *N*-Boc glycine in the presence of EDCI and HOBt in DMF was followed by treatment with trifluoroacetic acid to give aminoacetamide **12**. Hydroxyacetamide **13** was obtained in one step using the procedure described for **12**. The amino group of **58** was acylated with bromoacetyl bromide to yield bromoacetamide **18**. Bromide **18** was treated with potassium thioacetate to give thioacetate **15**, after which deacetylation of the thioacetate in the presence of K_2CO_3 in MeOH gave mercaptoacetamide **14**. The amine **58** was allowed to react with propargyl bromide in the presence of K_2CO_3 to give mono- and di-alkylated compounds **16** and **17**.

Compounds **6**−**9**, **11**, **19**−**21**, **24**−**32**, and **37** were prepared from another starting material, **59** (Scheme 2). The preparation of hydroxysulfonamide **6** was achieved via sulfonyl chloride **60**. Bromide **59** was converted to sulfonyl chloride **60** by sulfation and by a

Scheme 2[a]



[a] Reagents and conditions: (a) Na_2SO_3, EtOH, H_2O, reflux; (b) $SOCl_2$, DMF, toluene, reflux; (c) *O*-(2-tetrahydropyranyl)hydroxylamine, 4-(dimethylamino)pyridine, pyridine, CH_2Cl_2, rt; (d) 2N aq NaOH, EtOH, rt; (e) aniline, EDCI, HOBt, DMF, rt; (f) TFA, CH_2Cl_2, 60 °C; (g) LiOH·H_2O, EtOH, THF, H_2O, rt; (h) $(COCl)_2$, DMF, CH_2Cl_2, rt; (i) $ArNH_2$ (**63**), Et_3N, CH_2Cl_2, rt; (j) $PhB(OH)_2$, $Pd(PPh_3)_4$, $NaHCO_3$, 1-methyl-2-pyrrolidinone, H_2O, 80 °C; (k) AcSK, EtOH, rt; (l) 15% aq NaSMe, EtOH, rt; (m) *m*-chloroperoxybenzoic acid, CH_2Cl_2, rt.

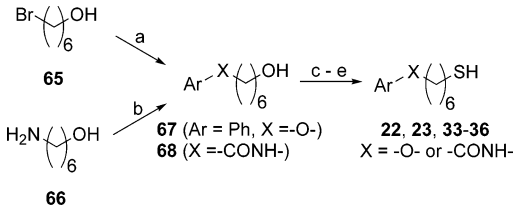
subsequent reaction with thionyl chloride. The sulfonyl chloride **60** was treated with *O*-THP hydroxylamine to give *O*-THP hydroxysulfonamide, after which hydrolysis of the ester under alkaline conditions and subsequent amide formation with aniline gave compound **61**. Removal of the THP group of compound **61** by treatment with trifluoroacetic acid gave hydroxysulfonamide **6**.

Compounds **7**−**9**, **11**, **19**−**21**, **24**−**32**, and **37** were synthesized from the corresponding acid chlorides **62** (**62a** ($n = 4$) and **62b** ($n = 5$) are commercially available) by the route shown in Scheme 2. **62c** ($n = 6$) was prepared from ester **59** by hydrolysis of the ethyl ester and a subsequent reaction with oxalyl chloride, and **62d** ($n = 7$) was obtained in the same way as **62c**. The amino group of aromatic amines **63** was acylated with an appropriate acid chloride **62** to give the amides **64**. Suzuki coupling[14] of bromobenzene **64a** with phenylboronic acid provided the biphenyl **64b**. Bromides **64** were treated with potassium thioacetate to give compound **8**, after which hydrolysis of the thioacetates under alkaline conditions gave the desired compounds **7**, **19**−**21**, **24**−**31**, and **32**, and disulfide **37** was obtained as a byproduct when thiol **7** was synthesized. Sulfide **9** was prepared by the alkylation of methylmercaptan with bromide **64c** (Ar = Ph, $n = 6$). Oxidation of **9** with 2 equiv of *m*-chloroperoxybenzoic acid provided the sulfone **11**.

Thiols **22**, **23**, **33**−**35**, and **36** were prepared from alcohol **65** or **66** by the procedure outlined in Scheme 3. Treatment of bromide **65** with phenol in the presence of K_2CO_3 gave ether **67**, and condensation of amine **66** with an appropriate aromatic carboxylic acid **69** afforded amides **68**. Alcohols **67** and **68** were converted to thiols **22**, **23**, **33**−**35**, and **36** in three steps by conversion of the alcohols to bromides, treatment of the bromides with potassium thioacetate, and hydrolysis of the resulting thioacetates.

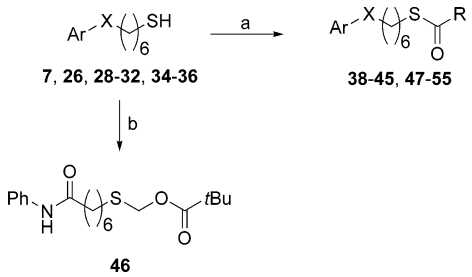
The preparation of *S*-chemically modified compounds **38**−**54**, and **55** is shown in Scheme 4. Thiols **7**, **26**, **28**−**32**, **34**, **35**, and **36** were coupled with the corresponding

Scheme 3[a]



[a] Reagents and conditions: (a) Phenol, K_2CO_3, DMF, 80 °C; (b) ArCOOH (**69**), EDCI, HOBt, DMF, rt; (c) CBr_4, PPh_3, CH_2Cl_2, 0 °C; (d) AcSK, EtOH, rt; (e) 2N aq NaOH, EtOH, THF, rt.

Scheme 4[a]



[a] Reagents and conditions: (a) RCOCl (**70**), 4-(dimethylamino)pyridine, pyridine, CH_2Cl_2, rt; (b) NaH, chloromethyl pivalate, DMF, 0 °C to room temperature.

acyl chloride **70** to give thioesters **38**−**45**, **47**−**54**, and **55**. Alkylation of thiol **7** with chloromethyl pivalate in the presence of sodium hydride in DMF afforded compound **46**.

Results and Discussion

Enzyme Assays. The compounds synthesized in this study were tested with an in vitro assay using a HeLa nuclear extract rich in HDAC activity. The results are summarized in Tables 1−3.

The IC_{50} values of SAHA and *o*-aminoanilide **1** were 0.28 μM and 120 μM, respectively (entries 1 and 2). α-Ketoamide **2** and *N*-formyl hydroxylamine **3** were reported previously to inhibit HDACs with an IC_{50} of 0.34 μM and 2.8 μM, respectively (entries 3 and 4).[11b,12]

The crystal structures of an archaebacterial HDAC homologue (HDAC-like protein, HDLP)/hydroxamates and HDAC8/hydroxamates complexes made it clear that the hydroxamic acid group coordinates the zinc ion in the active site through its CO and OH groups and also forms three hydrogen bonds between its CO, NH, and OH groups and Tyr 306, His 143, and His 142 (HDAC8 numbering), respectively.[7] From these data, hydroxyurea **4**, semicarbazide **5**, and hydroxysulfonamide **6** were synthesized and tested as HDAC inhibitors because it is possible for them to chelate zinc ion and form hydrogen bonds with Tyr and His like SAHA. Among these three compounds, hydroxyurea **4** and semicarbazide **5** showed anti-HDAC activity and the IC_{50} values were comparable to that of *o*-aminoanilide **1** (entries 5, 6, and 7). However, they were much less effective than SAHA.

Thiols seemed to be reasonable targets for hydroxamic acid replacements, because zinc ion is highly thiophilic and thiol derivatives have been reported to inhibit zinc-dependent enzymes such as angiotensin converting enzyme[15] and matrix metalloproteinases.[16] Furthermore, macrocyclic compounds bearing a disulfide group such as FK228 have been reported recently to inhibit

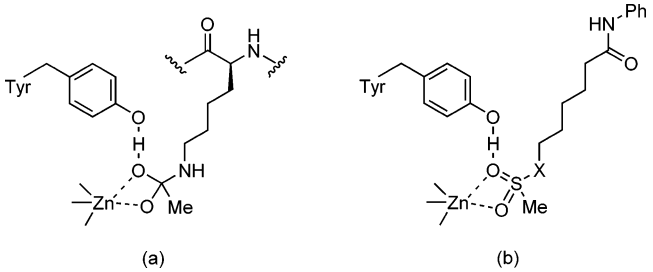


Figure 1. The transition state proposed for HDACs (a), and models for the binding of sulfone derivatives (b).

HDACs under reductive conditions.[17] Surprisingly, although the inhibitory ability of monodentate ZBGs such as thiol was thought to be less than that of bidentate ZBGs such as hydroxamate, *N*-formyl hydroxylamine, and hydrated electrophilic ketones,[16] the activity of thiol **7** was far greater than expected. A pronounced inhibitory effect (IC_{50} = 0.21 μM) was observed with thiol **7**, which was much more active than previously reported non-hydroxamates such as *o*-aminoanilide, *N*-formyl hydroxylamine, and trifluoromethyl ketone,[11a] and as potent as α-ketoamide **2** and SAHA (entry 8). To confirm that the thiol group plays an important role in anti-HDAC activity, thioacetate **8a** and methyl sulfide **9** were tested. As expected, thiol transformation into thioacetate and methyl sulfide led to an inhibitor that was about 30-fold less potent and a compound devoid of anti-HDAC activity, respectively (entries 9 and 10). These results suggest that thiolate anion generated under physiological conditions has an intimate involvement in the interaction with the zinc ion in the active site.

The crystal structures of the HDLP/hydroxamates and HDAC8/hydroxamates complexes have led to a solid understanding of not only the three-dimensional structure of the active site of HDACs but also the catalytic mechanism for the deacetylation of acetylated lysine substrate.[7] It has been proposed that the carbonyl oxygen of this substrate could bind the zinc, and the carbonyl could be attacked by a zinc-chelating water molecule (Figure 2a), which would result in the production of deacetylated lysine via a tetrahedral carbon-containing transition state (Figure 1a). On the basis of the proposed catalytic mechanism, we attempted to design non-hydroxamate HDAC inhibitors. First, we designed transition-state (TS) analogues. The TS of HDAC deacetylation was estimated to include a tetrahedral carbon (Figure 1a) as with other zinc proteases.[18] We focused attention on sulfone derivative TS analogues because it has been suggested that the sulfonamide moiety has strong similarity with the TS of amide bond hydrolysis, both from a steric and an electronic point of view.[19] Compounds **10** and **11**, in which a hydroxamic acid of SAHA is replaced by a sulfonamide and a sulfone, respectively, were designed and synthesized as TS analogues (Figure 1b). Of these two TS analogues, sulfone **11** showed anti-HDAC activity and the IC_{50} value was 230 μM (entries 11 and 12). However, sulfone **11** was approximately 820-fold less effective than SAHA.

Our next approach was based on the proposed deacetylation mechanism whereby a zinc-chelating water molecule activated by His142 and His 143 (HDAC8 numbering) makes a nucleophilic attack on the carbonyl carbon of an acetylated lysine substrate (Figure 2a).

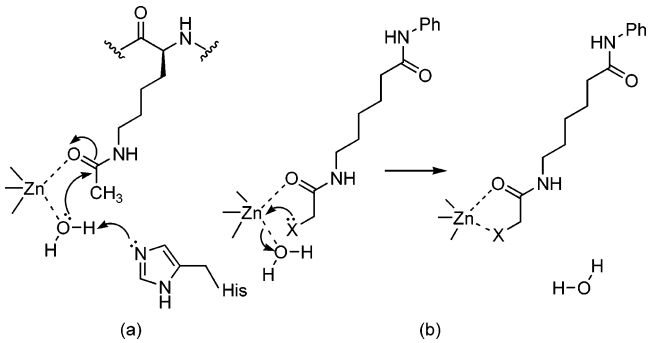
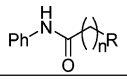
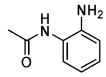


Figure 2. The mechanism proposed for the deacetylation of acetylated lysine substrate (a), and a model for the binding of heteroatom-containing substrate analogues to zinc ion (b).

Table 1. HDAC Inhibition Data for SAHA and SAHA-based Non-hydroxamtes[a]

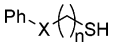


entry	compd	R	*n*	% inhbtn at 100 μM	IC_{50} (μM)
1	SAHA[b]	-CONHOH	6	100	0.28
2	**1**[c]		6	48	120
3	**2**	-COCONHMe	6	ND	0.34[d]
4	**3**		7	ND	2.8[e]
5	**4**	-NHCONHOH	5	58	80
6	**5**	-NHCONHNH$_2$	5	35	150
7	**6**	-SO$_2$NHOH	6	14	>100
8	**7**	-SH	6	100	0.21
9	**8a**	-SAc	6	85	7.1
10	**9**	-SMe	6	11	>100
11	**10**	-NHSO$_2$Me	5	10	7500
12	**11**	-SO$_2$Me	6	33	230
13	**12**[f]	-NHCOCH$_2$NH$_2$	5	6	>100
14	**13**	-NHCOCH$_2$OH	5	0	>100
15	**14**	-NHCOCH$_2$SH	5	99	0.39
16	**15**	-NHCOCH$_2$SAc	5	72	22
17	**16**		5	14	>100
18	**17**		5	0	>100
19	**18**	-NHCOCH$_2$Br	5	78	17

[a] Values are means of at least three experiments. [b] Prepared as described in ref 26. [c] Prepared as described in ref 9a. [d] Data taken from the literature (ref 11b). [e] Data taken from the literature (ref 12). [f] Trifluoroacetic acid salt. ND = No data.

With this mechanism, if the water molecule is forcibly removed from the zinc ion, the HDACs would supposedly be inhibited. We then designed and synthesized heteroatom-containing substrate analogues **12**, **13**, and **14**. These analogues would be recognized as substrates by HDACs and would be easily taken into the active site where they could force the water molecule off the zinc ion and the reactive site for the deacetylation by chelation of the heteroatom to the zinc ion, and might behave as HDAC inhibitors (Figure 2b). As shown in Table 1 (entries 13, 14, and 15), potent inhibition was observed with mercaptoacetamide **14**, while **12** and **13** did not possess HDAC inhibitory activities. Mercaptoacetamide **14** exhibited an IC_{50} of 0.39 μM, and its activity largely surpassed those of *o*-aminoanilide **1** and *N*-formyl hydroxylamine **3** and was comparable to those of α-ketoamide **2** and SAHA. As expected, thiol transformation into thioacetate (**15**) led to a 55-fold less potent inhibitor. This result suggests the ease of ionization of thiol is an important factor for HDAC inhibition like the case of thiol **7**.

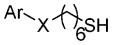
Table 2. Effect of Linker Variation on HDAC Inhibitory Activity of Thiols[a]

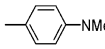
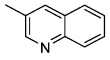
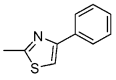
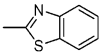
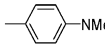
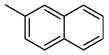
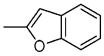
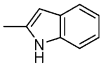


entry	compd	X	*n*	IC_{50} (μM)
1	**7**	-NHCO-	6	0.21
2	**19**	-NHCO-	7	1.5
3	**20**	-NHCO-	5	0.37
4	**21**	-NHCO-	4	6.2
5	**22**	-O-	6	11
6	**23**	-CONH-	6	0.36

[a] Values are means of at least three experiments.

Table 3. Effect of Aromatic Group Variation on HDAC Inhibitory Activity of Thiols[a]

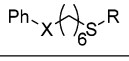


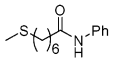
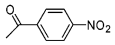
entry	compd	Ar	X	IC_{50} (μM)
1	**7**	-Ph	-NHCO-	0.21
2	**24**		-NHCO-	1.2
3	**25**		-NHCO-	1.1
4	**26**		-NHCO-	0.075
5	**27**		-NHCO-	0.62
6	**28**		-NHCO-	0.21
7	**29**		-NHCO-	0.11
8	**30**		-NHCO-	0.072
9	**31**		-NHCO-	0.17
10	**32**		-NHCO-	0.34
11	**23**	-Ph	-CONH-	0.36
12	**33**		-CONH-	0.61
13	**34**		-CONH-	0.085
14	**35**		-CONH-	0.079
15	**36**		-CONH-	0.10

[a] Values are means of at least three experiments.

We turned our attention to irreversible HDAC inhibitors. TPX B is an irreversible HDAC inhibitor,[20] and finding more specific and simpler irreversible HDAC inhibitors is useful for the isolation and cloning of an HDAC.[2] As described above, the crystal structures of the HDLP/hydroxamates and HDAC8/hydroxamates complexes revealed that the hydroxamic acid group forms three hydrogen bonds with Tyr 306, His 143, and His 142, and furthermore, zinc ion is coordinated by His 180, Asp 178, and Asp 267 (HDAC8 numbering). Since the phenol group of Tyr, the imidazole group of His, and the carboxyl group of Asp are able to react with electrophiles, we prepared analogues bearing propargyl

Table 4. Cell Growth Inhibition Data on NCI-H460 Cells for Compound **7** and Its *S*-Modified Prodrugs[a]



entry	compd	R	EC_{50} (μM)
1	**7**	-H	>50[b]
2	**37**		>50[c]
3	**8a**	-Ac	36
4	**38**	-COEt	28
5	**39**	-CO*n*-Pr	22
6	**40**	-CO*i*-Pr	20
7	**41**	-CO*t*-Bu	>50[d]
8	**42**		27
9	**43**		21
10	**44**	-Bz	25
11	**45**		24
12	**46**	-CH_2OCO*t*-Bu	25

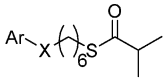
[a] Values are means of at least two experiments. [b] 34% inhibition at 50 μM. [c] 10% inhibition at 50 μM. [d] 42% inhibition at 50 μM.

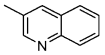
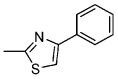
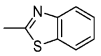
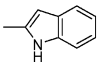
amino (**16**, **17**) and bromoacetamide (**18**) which could form covalent bonds with Tyr, His, and Asp of the enzyme, and evaluated their anti-HDAC activities. While propargyl amino compounds **16** and **17** did not possess HDAC inhibitory activities, more potent inhibition was observed with bromoacetamide **18** (entries 17, 18, and 19). Bromoacetamide **18** exhibited an IC_{50} of 17 μM and its activity was about 9-fold as strong as that of *o*-aminoanilide **1**, but much weaker than that of SAHA.

With the results shown in Table 1, we were encouraged to study further the structure−activity relationship (SAR) and structural optimization. We selected thiol **7** for further study.[21] First, we examined the effect of linker parts of thiol **7**. The results are summarized in Table 2. HDAC inhibition was distinctly dependent on chain length, with $n = 7$ (**19**) and $n = 4$ (**21**) resulting in less potent inhibitors. However, compound **20**, in which $n = 5$, showed essentially the same potency as compound **7**, in which $n = 6$ (entries 1−4). The similar SAR between thiols and hydroxamates, with $n = 6$ optimal,[22] indicates that thiols inhibit HDACs in a binding mode similar to that of hydroxamates. As for the group attaching the phenyl moiety, ether **22** displayed moderate activity, whereas the activity of the reversed amide **23** was maintained (entries 5 and 6).

Next, the aromatic group was examined (Table 3). In the amide-linked series (entries 1−10), 4-substituted phenyl compounds tended to decrease the potency. Specifically, compounds **24** (Ar = 4-NMe_2-Ph), **25** (Ar = 4-biphenyl), and **27** (Ar = 4-PhO-Ph) showed about a 3- to 6-fold decrease in potency when compared to the parent thiol **7** (entries 2, 3, and 5). On the other hand, compound **26**, in which a phenyl group was introduced at the 3-position of the phenyl group of **7**, showed 3-fold increased inhibitory activity (IC_{50} of 0.075 μM, entry 4). In addition, 3-phenoxy compound **28** was equipotent with compound **7** (entry 6). We investigated the effect of the replacement of the phenyl group of compound **7** with heteroaryl rings (entries 7, 8, 9, and 10). Changing the benzene ring to a 3-pyridine ring (**29**), 4-phenyl-2-thiazole ring (**31**), and 2-benzothiazole ring (**32**) sustained or slightly reduced the activity, whereas quinoline **30** had improved activity (IC_{50} of 0.072 μM), and turned out to be the most potent compound in this series. The reverse amide-linked series (entries 11−15) exhibited potencies similar to or greater than the parent thiol **23**, with the exception of **33** (Ar = 4-NMe_2-Ph), which resulted in a slightly less potent inhibitor. In particular, the reversed amides **34** with a naphthalene substituent and **35** with a benzofuran substituent exhibited about 3-fold increases in potency (IC_{50}s of 0.085 μM and 0.079 μM, respectively). As a result, IC_{50}s in the double-digit nanomolar range were observed with 3-biphenyl **26**, quinoline **30**, naphthalene **34**, and benzofuran **35**, which were approximately 3- to 4-fold more potent than SAHA.

Table 5. Cell Growth Inhibition Data on NCI-H460 Cells for Compound **40** and Its Derivatives[a]



entry	compd	Ar	X	EC_{50} (μM)
1	**40**	-Ph	-NHCO-	20
2	**47**		-NHCO-	2.8
3	**48**		-NHCO-	25
4	**49**		-NHCO-	2.9
5	**50**		-NHCO-	8.0
6	**51**		-NHCO-	2.1
7	**52**		-NHCO-	9.5
8	**53**		-CONH-	12
9	**54**		-CONH-	4.1
10	**55**		-CONH-	12

[a] Values are means of at least two experiments.

Table 6. Growth Inhibition of Various Cancer Cells Using SAHA and Compound **51**[a]

	cell	SAHA, EC_{50} (μM)	**51**, EC_{50} (μM)
MDA-MB-231	breast cancer	1.5	2.3
SNB-78	central nervous system	16	9.1
HCT116	colon cancer	0.58	3.0
NCI−H226	lung cancer	2.6	2.6
LOX-IMVI	melanoma	1.3	1.1
SK-OV-3	ovarian cancer	2.5	4.5
RXF-631L	renal cancer	2.0	2.4
St-4	stomach cancer	5.2	5.0
DU-145	prostate cancer	1.6	4.5
	mean	3.7	3.8

[a] Values are means of at least two experiments.

Cancer Cell Growth Inhibition Assay. To confirm the effectiveness of thiol-based HDAC inhibitors as anticancer drugs and tools for biological research, thiol **7** was initially tested in a cancer cell growth inhibition

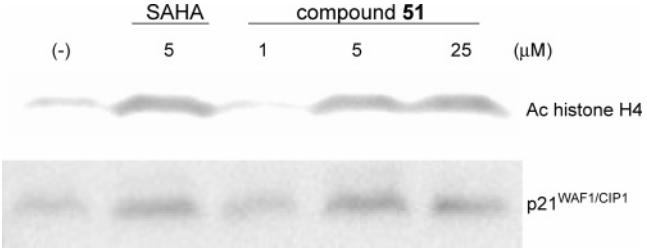


Figure 3. Western blot analysis of histone hyperacetylation and p21$^{WAF1/CIP1}$ induction in HCT 116 cells produced by compound **51** and by reference compound SAHA.

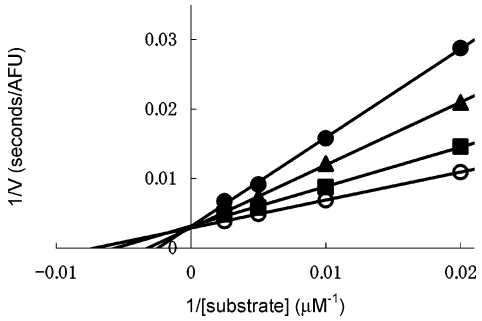


Figure 4. Reciprocal rate vs reciprocal acetylated lysine substrate concentration in the presence of 0.3 (●), 0.1 (▲), 0.03 (■), and 0 (○) μM of **7**.

assay using human lung cancer NCI-H460 cells against which it was found to be only weakly potent, although **7** was highly active in an enzymatic assay (Table 4, entry 1). The reason for the weak activity of thiol **7** is unclear, but it is reasonable to assume that it was due to poor membrane permeability resulting from the highly polar character of this compound, and that a transient masking of the sulfhydryl group could improve its permeability and its ability to inhibit cancer cell growth. Therefore, we investigated the possibility of improving the inhibition using the prodrug approach. In the search for a suitable prodrug of thiols, disulfides seemed to be attractive targets, because it has been reported that the disulfide bond of macrocyclic compounds bearing a disulfide group such as FK228 is reduced in the cellular environment, releasing the free thiol analogue as the active species.[17] However, contrary to our expectation, disulfide **37** failed to exhibit a growth inhibitory effect on NCI-H460 cells (entry 2). Next, we examined the activity of acetyl compound **8a**. Acetyl compound **8a** proved to be relatively potent compared with thiol **7** and disulfide **37** (EC_{50} of 36 μM) (entry 3). This result suggests that **8a** permeates the cell membrane more efficiently than thiol **7**, and is converted to thiol **7** by enzymatic hydrolysis within the cell.[23] Encouraged by this finding, we prepared other *S*-acyl compounds (**38**−**45**) and evaluated their activities (entries 4−11). This series of compounds exhibited greater potency than acetyl compound **8a**, except for pivaloyl compound **41**, which was a less potent inhibitor. In particular, isobutyryl compound **40** showed about a 2-fold increase in activity when compared to acetyl compound **8a** (EC_{50} of 20 μM). The compound bearing a (pivaloyloxy)methyl group[24] (**46**) was slightly less active than isobutyryl compound **40** (entry 12).

With the results shown in Table 4, a selected set of active compounds from the enzymatic assay was *S*-isobutyrylated and tested as cancer cell growth inhibitors (Table 5). Much to our satisfaction, changing the phenyl group of compound **40** to other aromatic groups led to positive results. Isobutyryl analogues **47**−**55** were generally more potent than the parent compound **40**; the sole exception is **48** (Ar = 3-OPh-Ph) which was slightly less active than compound **40** (entry 3). Notably, 3-biphenyl (**47**), 3-pyridinyl (**49**), and 4-phenyl-2-thiazolyl (**51**) analogues showed strong activity in inhibiting the growth of NCI-H460 cells, with EC_{50}s of 2−3 μM. Furthermore, we evaluated cancer cell growth inhibition by SAHA and **51**, the most potent compound in this study, against nine other human cancer cell lines

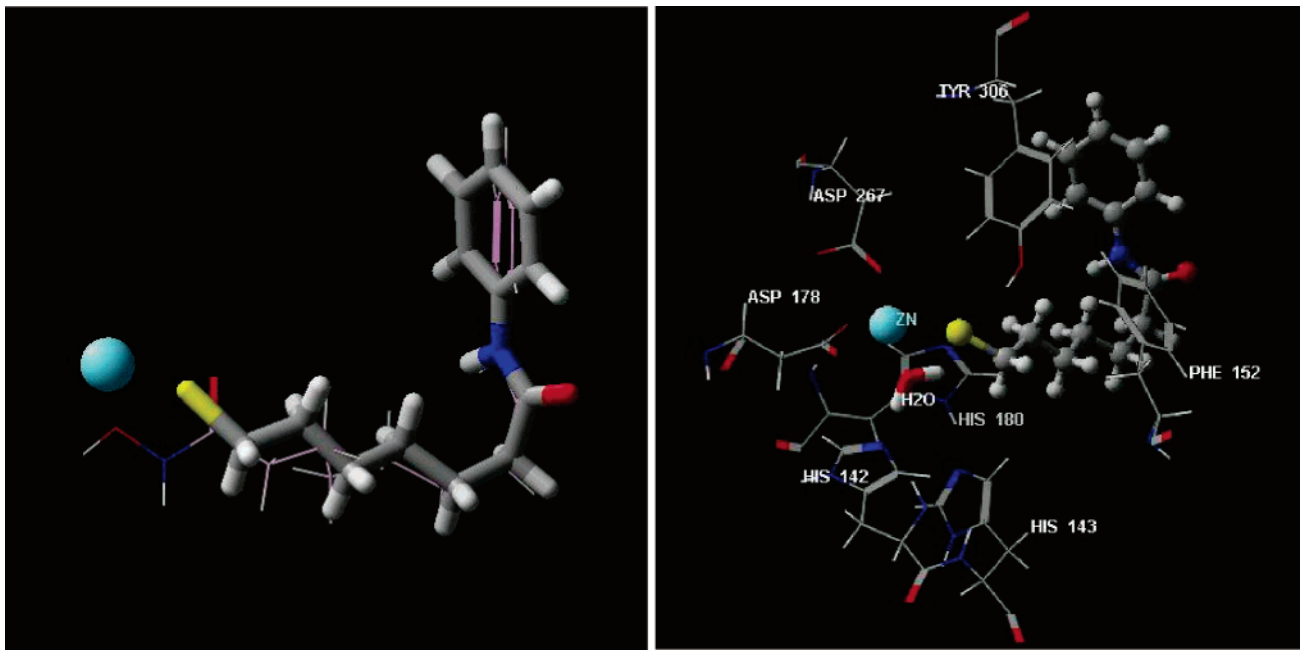


Figure 5. Superposition of the low energy conformations of **7** (tube) and SAHA (wire) (left). The HDAC8 pocket is not shown for the sake of clarity. View of the conformation of **7** (ball-and-stick) docked in the HDAC8 catalytic core (right). Residues around the zinc ion and a water molecule are displayed as wires and tubes, respectively.

(Table 6). Compound **51** exerted potent growth inhibition against various human cancer cells, with EC_{50} values ranging from 1 to 10 μM, and these inhibitory activities were comparable to those of SAHA (average EC_{50} of **51** 3.8 μM, SAHA 3.7 μM) which is currently being evaluated in clinical trials for use in the treatment of cancer.

By Western blot analysis, cancer cell growth inhibition with compound **51** was verified to be the result of inhibition of HDACs (Figure 3). Treatment of HCT 116 cells with compound **51** gave rise to elevated and dose-dependent levels of acetylated histone H4 and $p21^{WAF1/CIP1}$.

Inhibitory Mechanism Study. Since the results of cancer cell growth inhibition and Western blot analysis have suggested that thiols generated from *S*-acyl prodrugs by enzymatic hydrolysis within the cell inhibit intracellular HDACs, we next studied the mechanism by which thiols inhibit HDACs in greater detail. Although the sulfhydryl group of thiol derivatives was designed as a ZBG, it is possible that thiols inhibit HDACs by forming a covalent disulfide bond with cysteine residues on these enzymes. We examined this possibility using a double reciprocal plot of 1/V versus 1/[substrate] at varying concentrations of inhibitor **7** (Figure 4), and the data from this study established that thiol **7** engages in competitive inhibition versus acetylated lysine substrate, with an inhibition constant (K_i) of 0.11 μM. Since cysteine is not a component in the construction of the active site of HDACs, the sulfhydryl group of **7** likely interacts with the zinc in the active site.

Binding Mode Study. Since thiol **7** proved to be a competitive inhibitor and to act within the active center of HDACs, we studied its binding mode within this site. The low energy conformations of **7** and SAHA were calculated when docked in the model based on the crystal structure of HDAC8 (PDB code 1T64, 1T67, 1T69, and 1VKG) using Macromodel 8.1 software.[25] The anilide group and alkyl chain of **7** and SAHA were essentially superimposed in the binding pocket, and the binding mode of **7** was found to be similar to that of SAHA (Figure 5, left). An inspection of the HDAC8/**7** complex shows that the sulfur atom of **7** was located 2.35 Å from the zinc ion, 2.24 Å from the OH group of Tyr 306, and 2.66 Å from a water molecule which forms a hydrogen bond with the imidazole group of His142 (Figure 5, right). This suggests that thiols strongly inhibit HDACs by interacting directly with zinc ion, Tyr 306, and His 142 via a water molecule.

Conclusion

We have designed and prepared a series of SAHA-based compounds as (i) hydroxamic acid mimics by structure-based drug design (compounds **4**−**6**), (ii) thiol-based analogues (compounds **7**−**9**), (iii) transition-state analogues (compounds **10** and **11**), (iv) heteroatom-containing substrate analogues by mechanism-based drug design (compounds **12**−**15**), and (v) irreversible inhibition-oriented compounds (compounds **16**−**18**), and evaluated their inhibitory effect on HDACs. In this series, thiol **7** and mercaptoacetamide **14** were found to be much more potent HDAC inhibitors than previously reported non-hydroxamates, and as potent as α-ketoamide **2** and SAHA. At present, thiol is one of the most active ZBG among small-molecule HDAC inhibitors. Optimization of thiol derivatives led to the identification of inhibitors more effective than SAHA (compounds **26**, **30**, **34**, and **35**). We have also identified a potent cancer cell growth inhibitor, compound **51**, by the prodrug formation of thiol-based HDAC inhibitors. Thiol **7** exhibits strong competitive inhibition of an acetylated lysine substrate, and molecular modeling suggests that the thiol interacts with zinc, Tyr 306, and His 142 (HDAC8 numbering) in the active site.

In conclusion, we have identified several new lead structures including thiol, from which more potent HDAC inhibitors can be developed. As far as we could determine, this is the first systematic study of ZBGs for HDAC inhibitors. We believe that the findings of this study should be of value in future studies for the development of ideal anticancer drugs and tools for biological research such as HDAC isozyme-selective inhibitors.

Experimental Section

Chemistry. Melting points were determined using a Yanagimoto micro melting point apparatus or a Büchi 545 melting point apparatus and were left uncorrected. Proton nuclear magnetic resonance spectra (^{1}H NMR) were recorded on a JEOL JNM-LA400 or JEOL JNM-LA500 spectrometer in solvent as indicated. Chemical shifts (δ) are reported in parts per million relative to the internal standard tetramethylsilane. Elemental analysis was performed with a Yanaco CHN CORDER NT-5 analyzer, and all values were within ±0.4% of the calculated values. High-resolution mass spectra (HRMS) were recorded on a JEOL JMS−SX102A mass spectrometer. GC-MS analyses were performed on a Shimadzu GCMS-QP2010. Reagents and solvents were purchased from Aldrich, Tokyo Kasei Kogyo, Wako Pure Chemical Industries, and Kanto Kagaku and used without purification. Flash column chromatography was performed using silica gel 60 (particle size 0.046−0.063 mm) supplied by Merck.

6-(3-Hydroxyureido)hexanoic Acid Phenylamide (4). Step 1: Preparation of 6-Phenylcarbamoylhexanoic Acid (57). A mixture of aniline (5.80 g, 62.3 mmol) and pimeric acid (**56**, 10.0 g, 62.4 mmol) was stirred at 180 °C for 1 h. After cooling, the mixture was diluted with AcOEt-THF and the slurry was filtered. The filtrate was washed with saturated aqueous $NaHCO_3$, and the aqueous layer was acidified with concentrated HCl. The precipitated crystals were collected by filtration to give 7.11 g (49%) of **57** as a white solid: ^{1}H NMR (DMSO-d_6, 400 MHz, δ; ppm) 11.97 (1H, broad s), 9.83 (1H, s), 7.58 (2H, d, J = 7.8 Hz), 7.27 (2H, t, J = 7.9 Hz), 7.01 (1H, t, J = 7.4 Hz), 2.67 (2H, t, J = 7.4 Hz), 2.21 (2H, t, J = 7.3 Hz), 1.62−1.49 (4H, m), 1.34−1.27 (2H, m).

Steps 2 and 3: Preparation of 6-(3-Hydroxyureido)-hexanoic Acid Phenylamide (4). To a suspension of **57** (958 mg, 4.07 mmol) obtained above and triethylamine (744 mg, 7.35 mmol) in toluene (10 mL) was added diphenylphosphoryl azide (1.75 g, 6.34 mmol), and the mixture was heated at reflux temperature for 1h. Next, *O*-(2-tetrahydropyranyl)hydroxylamine (380 mg, 3.11 mmol) was added, and the reaction mixture was stirred at reflux temperature for 18h. It was then concentrated in vacuo, and the residue was dissolved in AcOEt. The AcOEt solution was washed with water, saturated aqueous $NaHCO_3$, and brine and was dried over Na_2SO_4. Filtration and concentration in vacuo and purification by silica gel flash chromatography (*n*-hexane/AcOEt = 1/2) gave 988 mg (69%) of the *O*-(2-tetrahydropyranyl)hydroxyurea as a white solid: ^{1}H NMR ($CDCl_3$, 500 MHz, δ; ppm) 7.53 (2H, d, J = 7.9 Hz), 7.32 (2H, t, J = 7.8 Hz), 7.26 (1H, broad s), 7.10 (1H, t, J = 7 Hz), 7.05 (1H, broad s), 6.06 (1H, broad s), 4.75 (1H, d, J = 3.6 Hz), 3.93 (1H, m), 3.57 (1H, m), 3.33−3.26 (2H, m), 2.38 (2H, t, J = 7.5 Hz), 1.82−1.77 (4H, m), 1.61−1.55 (6H, m), 1.44 (2H, quintet, J = 7.3 Hz).

To a solution of the *O*-(2-tetrahydropyranyl)hydroxyurea (185 mg, 0.53 mmol) obtained above in MeOH (2 mL) was added 4-toluenesulfonic acid monohydrate (15 mg, 0.079 mmol). The solution was stirred overnight at room temperature, and the precipitated crystals were collected by filtration to give 46 mg (32%) of **4** as a white solid. The solid was recrystallized from MeOH−AcOEt and collected by filtration to give 34 mg of **4** as a colorless crystal: mp 148−149 °C; ^{1}H NMR (DMSO-d_6, 500 MHz, δ; ppm) 9.93 (1H, s), 8.58 (1H, s), 8.29 (1H, s), 7.65 (2H, d, J = 8 Hz), 7.35 (2H, t, J = 7.9 Hz), 7.08 (1H, t, J = 7.3 Hz), 6.75 (1H, t, J = 6 Hz), 3.10 (2H, q, J = 6.7 Hz), 2.36 (2H, t, J = 7.5 Hz), 1.65 (2H, quintet, J = 7.5 Hz), 1.50 (2H, quintet, J = 7.2 Hz), 1.34 (2H, quintet, J = 7.6 Hz); Anal. ($C_{13}H_{19}N_3O_3$) C, H, N.

6-(3-Aminoureido)hexanoic Acid Phenylamide (5). Compound **5** was prepared from **57** obtained above by using the procedure described for **4** (step 2) in 52% yield. In this case, hydrazine monohydrate was used instead of *O*-(2-tetrahydropyranyl)hydroxylamine: mp 146−147 °C; ^{1}H NMR (DMSO-d_6, 400 MHz, δ; ppm) 9.83 (1H, s), 7.58 (2H, d, J = 7.8 Hz), 7.27 (2H, t, J = 7.9 Hz), 7.01 (1H, t, J = 7.3 Hz), 6.83 (1H, broad s), 6.28 (1H, broad s), 4.03 (2H, broad s), 3.01 (2H, q, J = 6.7 Hz), 2.29 (2H, t, J = 7.4 Hz), 1.60−1.57 (2H, m), 1.40−1.38 (2H, m), 1.32−1.28 (2H, m); MS (EI) *m/z*: 264 (M^+); Anal. ($C_{13}H_{20}N_4O_2$) C, H, N.

6-Methanesulfonylaminohexanoic Acid Phenylamide (10). Steps 1 and 2: Preparation of 6-Aminohexanoic Acid Phenylamide (58). To a suspension of **57** (1.11 g, 4.73 mmol) obtained above and triethylamine (699 mg, 6.90 mmol) in benzene (3 mL) was added diphenylphosphoryl azide (1.83 g, 6.64 mmol), and the mixture was heated at reflux temperature for 1 h. Next, benzyl alcohol (1.20 mL, 11.6 mmol) was added, and the reaction mixture was stirred at reflux temperature for 24 h. It was then concentrated in vacuo and the residue was dissolved in AcOEt. The AcOEt solution was washed with 0.4 N aqueous HCl, water, saturated aqueous $NaHCO_3$, and brine and was dried over Na_2SO_4. Filtration and concentration in vacuo and purification by recrystallization from $CHCl_3$−*n*-hexane gave 1.01 g (63%) of (6-phenylcarbamoylpentyl)carbamic acid benzyl ester as a colorless needle: ^{1}H NMR (DMSO-d_6, 400 MHz, δ; ppm) 9.81 (1H, s), 7.57 (2H, d, J = 7.8 Hz), 7.37−7.22 (8H, m), 7.00 (1H, t, J = 7.4 Hz), 4.99 (2H, s), 2.99 (2H, q, J = 6.5 Hz), 2.28 (2H, t, J = 7.4 Hz), 1.58 (2H, quintet, J = 7.6 Hz), 1.43 (2H, quintet, J = 7.1 Hz), 1.32 (2H, quintet, J = 7.8 Hz); MS (EI) *m/z*: 340 (M^+).

A solution of (6-phenylcarbamoylpentyl)carbamic acid benzyl ester (1.00 g, 2.95 mmol) obtained above in MeOH (50 mL) was stirred under H_2 (atmospheric pressure) in the presence of 5% Pd/C (106 mg) at room temperature for 7 h. The catalyst was removed by filtration through Celite, and the filtrate was concentrated in vacuo. The residue was purified by silica gel flash chromatography ($CHCl_3$/MeOH/*i*PrNH$_2$ = 19/1/1) to give 584 mg (96%) of **58** as a white solid: ^{1}H NMR (DMSO-d_6, 400 MHz, δ; ppm) 9.83 (1H, s), 7.58 (2H, d, J = 7.6 Hz), 7.27 (2H, t, J = 7.9 Hz), 7.01 (1H, t, J = 7.3 Hz), 2.55 (2H, m), 2.29 (2H, t, J = 7.4 Hz), 1.59 (2H, quintet, J = 7.4 Hz), 1.37−1.30 (4H, m).

Step 3: Preparation of 6-Methanesulfonylaminohexanoic Acid Phenylamide (10). To a solution of **58** (500 mg, 2.06 mmol) obtained above in pyridine (5 mL) was added methanesulfonyl chloride (160 μL, 2.07 mmol) dropwise with cooling in an ice−water bath. The solution was stirred for 30 min at room temperature. The mixture was concentrated and diluted with AcOEt. The solution was washed with 2 N aqueous HCl, water, and brine and was dried over Na_2SO_4. Filtration and concentration in vacuo and purification by silica gel flash chromatography (*n*-hexane/AcOEt = 1/3) gave 418 mg (71%) of **10** as a crude solid. The solid was recrystallized from AcOEt to give **10** (214 mg) as colorless crystals: mp 136−137 °C; ^{1}H NMR (DMSO-d_6, 500 MHz, δ; ppm) 9.85 (1H, s), 7.58 (2H, d, J = 7.7 Hz), 7.28 (2H, t, J = 7.4 Hz), 7.01 (1H, t, J = 7.4 Hz), 6.93 (1H, t, J = 6.5 Hz), 2.92 (2H, q, J = 6.5 Hz), 2.87 (3H, s), 2.30 (2H, t, J = 7.6 Hz), 1.59 (2H, quintet, J = 7.6 Hz), 1.59 (2H, quintet, J = 7.6 Hz), 1.48 (2H, quintet, J = 7.4 Hz), 1.33 (2H, quintet, J = 7.4 Hz); MS (EI) *m/z*: 284 (M^+); Anal. ($C_{13}H_{20}N_2O_3S$) C, H, N.

6-(2-Hydroxyacetylamino)hexanoic Acid Phenylamide (13). To a solution of **58** (198 mg, 0.96 mmol) and glycolic acid (81 mg, 1.07 mmol) in DMF (6 mL) were added 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide hydrochloride (254 mg, 1.32 mmol) and 1-hydroxy-1*H*-benzotriazole monohydrate (244 mg, 1.59 mmol), and the mixture was stirred overnight at room temperature. The reaction mixture was poured into water and extracted with AcOEt. The AcOEt layer was separated, washed with saturated aqueous $NaHCO_3$ and brine, and dried over Na_2SO_4. Filtration and concentration in vacuo gave 251 mg (99%) of **13** as a crude solid. The solid was recrystallized from AcOEt to give 155 mg of **13** as a colorless crystal: mp 109−113 °C; ^{1}H NMR (DMSO-d_6, 500 MHz, δ; ppm) 9.92 (1H, s), 7.79 (1H, broad s), 7.65 (2H, d, J = 7.6 Hz), 7.35 (2H, t, J = 7.9 Hz), 7.08 (1H, t, J = 7.3 Hz), 5.51 (1H, t, J = 5.8 Hz), 3.84 (2H, d, J = 5.8 Hz), 3.16 (2H, q, J = 6.8 Hz), 2.36 (2H, t, J = 7.5 Hz), 1.65 (2H, quintet, J = 7.5 Hz), 1.51 (2H, quintet, J = 7.3 Hz), 1.35 (2H, quintet, J = 7.9 Hz); MS (EI) *m/z*: 264 (M^+); Anal. ($C_{14}H_{20}N_2O_3$) C, H, N.

6-(2-Aminoacetylamino)hexanoic Acid Phenylamide Trifluoroacetic Acid Salt (12·TFA). Step 1: Preparation of [(5-Phenylcarbamoylpentylcarbamoyl)methyl]carbamic Acid *tert*-Butyl Ester. This compound was prepared from **58** and *N*-(*tert*-butoxycarbonyl)glycine using the procedure described for **13** in 70% yield: ^{1}H NMR ($CDCl_3$, 400 MHz, δ; ppm) 7.53 (2H, d, J = 7.8 Hz), 7.34 (2H, t, J = 7.6 Hz), 7.10 (1H, t, J = 7.6 Hz), 6.14 (1H, broad s), 5.07 (1H, broad s), 3.75 (2H, d, J = 6 Hz), 3.30 (2H, q, J = 6.5 Hz), 2.37 (2H, t, J = 7.4 Hz), 1.76 (2H, quintet, J = 7.4 Hz), 1.58−1.26 (13H, m).

Step 2: Preparation of 6-(2-Aminoacetylamino)hexanoic Acid Phenylamide Trifluoroacetic Acid Salt (12·TFA). To a solution of [(5-phenylcarbamoylpentylcarbamoyl)-methyl]carbamic acid *tert*-butyl ester (147 mg, 0.40 mmol) obtained above in $CHCl_3$ (4 mL) was added trifluoroacetic acid (1 mL), and the mixture was stirred overnight at room temperature. The reaction mixture was concentrated in vacuo, and the residue was triturated in diethyl ether to give 131 mg (84%) of **12**·TFA as a white solid. The solid was recrystallized from AcOEt−MeOH to give 120 mg of **12**·TFA as colorless crystals: mp 149−151 °C; ^{1}H NMR (DMSO-d_6, 500 MHz, δ; ppm) 10.00 (1H, s), 8.43 (1H, t, J = 5.2 Hz), 8.10 (3H, broad s), 7.71 (2H, d, J = 8.2 Hz), 7.41 (2H, t, J = 7.9 Hz), 7.14 (1H, t, J = 7.3 Hz), 3.25 (2H, q, J = 6.4 Hz), 2.43 (2H, t, J = 7.3 Hz), 1.72 (2H, quintet, J = 7.5 Hz), 1.58 (2H, quintet, J = 7.2 Hz), 1.44 (2H, quintet, J = 7.5 Hz); Anal. ($C_{14}H_{21}N_3O_2$·TFA·1/10H_2O) C, H, N.

6-(2-Bromoacetylamino)hexanoic Acid Phenylamide (18). To a solution of **58** (70 mg, 0.340 mmol) and triethylamine (0.40 mL, 2.88 mmol) in THF (2 mL) was added a solution of bromoacetyl bromide (319 mg, 1.58 mmol) in THF (1 mL) dropwise with cooling in an ice−water bath. The mixture was stirred at room temperature for 30 min. The reaction mixture was diluted with $CHCl_3$, washed with aqueous saturated $NaHCO_3$, water, and brine, and dried over Na_2SO_4. Filtration and concentration in vacuo and purification by silica gel flash chromatography ($CHCl_3$/MeOH = 150/1) gave 25 mg (23%) of **18** as a white solid: ^{1}H NMR ($CDCl_3$, 400 MHz, δ; ppm) 7.52 (2H, d, J = 8.1 Hz), 7.32 (2H, t, J = 7.9 Hz), 7.19 (1H, broad s), 7.10 (1H, t, J = 7.6 Hz), 6.56 (1H, broad s), 3.87 (2H, s), 3.32 (2H, q, J = 6.6 Hz), 2.38 (2H, t, J = 7.3 Hz), 1.80−1.76 (2H, m), 1.63−1.59 (2H, m), 1.46−1.44 (2H, m); MS (EI) *m/z*: 326 (M^+); Anal. ($C_{14}H_{19}BrN_2O_2$) C, H, N.

Thioacetic acid *S*-[(6-Phenylcarbamoylpentylcarbamoyl)methyl] Ester (15). To a suspension of **18** (187 mg, 0.57 mmol) obtained above in EtOH (2 mL) was added potassium thioacetate (236 mg, 2.07 mmol), and the mixture was stirred at room temperature for 16 h. The reaction mixture was diluted with AcOEt and THF, washed with water and brine, and dried over Na_2SO_4. Filtration and concentration in vacuo and purification by silica gel flash chromatography (*n*-hexane/AcOEt = 1/1) gave 163 mg (89%) of **15** as a white solid. The solid was recrystallized from *n*-hexane−AcOEt to give 48 mg

of **15** as a colorless crystal: mp 130−133 °C; ^{1}H NMR ($CDCl_3$, 400 MHz, δ; ppm) 7.53 (2H, d, J = 7.3 Hz), 7.32 (2H, t, J = 7.9 Hz), 7.25 (1H, broad s), 7.10 (1H, t, J = 7.2 Hz), 6.25 (1H, broad s), 3.51 (2H, s), 3.25 (2H, q, J = 6.6 Hz), 2.39 (3H, s), 2.37 (2H, t, J = 7.4 Hz), 1.75 (2H, quintet, J = 7.6 Hz), 1.55 (2H, quintet, J = 7.1 Hz), 1.39 (2H, quintet, J = 7.3 Hz); MS (EI) *m/z*: 322 (M^+); Anal. ($C_{16}H_{22}N_2O_3S$) C, H, N.

6-(2-Mercaptoacetylamino)hexanoic Acid Phenylamide (14). To a solution of **15** (190 mg, 0.59 mmol) obtained above in MeOH (5 mL) was added K_2CO_3 (141 mg, 1.02 mmol), and the mixture was stirred at room temperature for 1 h. The reaction mixture was diluted with AcOEt and THF, washed with water and brine, and dried over Na_2SO_4. Filtration and concentration in vacuo and purification by silica gel flash chromatography ($CHCl_3$/MeOH = 20/1) gave 103 mg (62%) of **14** as a white solid. The solid was recrystallized from $CHCl_3$−MeOH to give 38 mg of **14** as a colorless crystal: mp 171−173 °C; ^{1}H NMR (DMSO-d_6, 500 MHz, δ; ppm) 9.84 (1H, s), 8.08 (1H, broad s), 7.57 (2H, d, J = 8.2 Hz), 7.27 (2H, t, J = 7.9 Hz), 7.00 (1H, t, J = 7.3 Hz), 3.90 (1H, s), 3.44 (2H, s), 3.07 (2H, q, J = 6.5 Hz), 2.28 (2H, t, J = 7.5 Hz), 1.59 (2H, quintet, J = 7.3 Hz), 1.45 (2H, quintet, J = 7 Hz), 1.31 (2H, quintet, J = 7.5 Hz); MS (EI) *m/z*: 280 (M^+); Anal. ($C_{14}H_{20}N_2O_2S$) C, H, N.

6-(2-Propynylamino)hexanoic Acid Phenylamide Hydrochloride Salt (16·HCl) and 6-(2-Dipropynylamino)-hexanoic Acid Phenylamide (17). To a solution of **58** (230 mg, 1.12 mmol) obtained above and K_2CO_3 (39 mg, 0.28 mmol) in MeOH (1 mL) was added propargyl bromide (38 mg, 0.32 mmol), and the mixture was stirred overnight at room temperature. The reaction mixture was concentrated in vacuo, and the residue was purified by silica gel flash chromatography ($CHCl_3$/MeOH = 15/1) to give 40 mg (51%) of **16** as a pale yellow oil and 12 mg (23%) of **17** as a pale yellow solid. To a solution of **16** in MeOH was added 1 N aqueous HCl (0.5 mL), and the solution was concentrated in vacuo. The residue was recrystallized from MeOH−AcOEt to give 16 mg of **16**·HCl as colorless needles: mp 161−165 °C; ^{1}H NMR (DMSO-d_6, 400 MHz, δ; ppm) 9.91 (1H, broad s), 9.12 (2H, broad s), 7.59 (2H, d, J = 7.6 Hz), 7.28 (2H, t, J = 7.9 Hz), 7.01 (1H, t, J = 7.3 Hz), 3.89 (2H, d, J = 3.4 Hz), 3.70 (1H, t, J = 2.6 Hz), 2.94 (2H, t, J = 7.8 Hz), 2.32 (2H, t, J = 7.3 Hz), 1.64−1.56 (4H, m), 1.36−1.25 (2H, m); MS (EI) *m/z*: 244 (M−-HCl); Anal. ($C_{15}H_{20}N_2O$·HCl·1/8H_2O) C, H, N.

The crude solid of **17** was recrystallized from $CHCl_3$−*n*-hexane to give 12 mg of **17** as colorless needles: mp 56−57 °C; ^{1}H NMR ($CDCl_3$, 400 MHz, δ; ppm) 7.51 (2H, d, J = 8.3 Hz), 7.32 (2H, t, J = 7.8 Hz), 7.11−7.10 (2H, m), 3.43 (4H, d, J = 2.4 Hz), 2.55 (2H, t, J = 7.3 Hz), 2.37 (2H, t, J = 7.4 Hz), 2.22 (2H, t, J = 2.3 Hz), 1.79−1.75 (2H, m), 1.54−1.52 (2H, m), 1.45−1.43 (2H, m); MS (EI) *m/z*: 281 (M^+); Anal. ($C_{18}H_{22}N_2O$) C, H, N.

7-Hydroxysulfamoylheptanoic Acid Phenylamide (6). Steps 1 and 2: Preparation of 7-Chlorosulfonylheptanoic Acid Ethyl Ester (60). To an aqueous solution (7 mL) of anhydrous sodium sulfite (2.03 g, 16.1 mmol) was added a solution of 7-bromoheptanoic acid ethyl ester (**59**, 2.0 g, 8.43 mmol) in EtOH (5 mL), and the solution was boiled under reflux with stirring for 2 h. The solution was evaporated to dryness, and the solid was dried in vacuo at 60 °C. This white solid was placed in a flask, toluene (30 mL) was added followed by a catalytic amount of DMF, and then thionyl chloride (6.2 mL, 85.0 mmol) was added dropwise. The mixture was boiled under reflux with stirring for 5 h, diluted with AcOEt, washed with aqueous saturated cold water and brine, and dried over $MgSO_4$. Filtration and concentration in vacuo and purification by silica gel flash chromatography (*n*-hexane/AcOEt = 4/1) gave 2.02 g (93%) of **60**: ^{1}H NMR ($CDCl_3$, 400 MHz, δ; ppm) 4.13 (2H, q, J = 7.1 Hz), 3.66 (2H, t, J = 7.8 Hz), 2.31 (2H, t, J = 7.3 Hz), 2.06 (2H, quintet, J = 7.8 Hz), 1.66 (2H, quintet, J = 7.3 Hz), 1.53 (2H, quintet, J = 7.8 Hz), 1.41 (2H, quintet, J = 7.1 Hz), 1.26 (2H, quintet, J = 7.1 Hz).

Steps 3, 4, and 5: Preparation of 7-(2-Tetrahydropyranyloxysulfamoyl)heptanoic Acid Phenylamide (61). To a mixture of *O*-(2-tetrahydropyranyl)hydroxylamine (251 mg, 2.14 mmol), a catalytic amount of 4-(dimethylamino)pyridine, pyridine (1 mL), and CH_2Cl_2 (10 mL) was added a solution of **60** (500 mg, 1.95 mmol) obtained above in CH_2Cl_2 (10 mL), and the mixture was stirred at room temperature for 5 h. The reaction mixture was poured into water and extracted with AcOEt. The AcOEt layer was separated, washed with water, saturated aqueous $NaHCO_3$ and brine, and dried over Na_2SO_4. Filtration and concentration in vacuo and purification by silica gel flash chromatography (*n*-hexane/AcOEt = 2/1) gave 618 mg (94%) of the sulfonamide as a crude oil.

To a solution of the sulfonamide(615 mg, 1.82 mmol) obtained above in EtOH (3 mL) was added 2 N aqueous NaOH (3.0 mL, 6.0 mmol). The mixture was stirred overnight at room temperature. The solvent was removed by evaporation in vacuo, and water was added to the residue. The mixture was neutralized with 2 N aqueous HCl (3.0 mL, 6.0 mmol) with cooling in an ice−water bath, and the mixture was extracted with AcOEt. The AcOEt layer was separated, washed with water and brine, and dried over Na_2SO_4. Filtration and concentration in vacuo gave 482 mg (86%) of the carboxylic acid as a white solid: ^{1}H NMR ($CDCl_3$, 400 MHz, δ; ppm) 7.40 (1H, broad s), 5.08 (1H, m), 3.93 (1H, m), 3.66 (1H, m), 3.21 (2H, m), 2.37 (2H, t, J = 7.3 Hz), 1.90−1.35 (14H, m).

Compound **61** was prepared from the carboxylic acid obtained above and aniline using the procedure described for **13** in 88% yield: ^{1}H NMR (DMSO-d_6, 400 MHz, δ; ppm) 10.04 (1H, broad s), 9.85 (1H, broad s), 7.58 (2H, d, J = 8 Hz), 7.28 (2H, t, J = 7.8 Hz), 7.01 (1H, t, J = 7.6 Hz), 4.88 (1H, m), 3.81 (1H, m), 3.52 (1H, m), 3.19−3.09 (2H, m), 2.30 (2H, t, J = 7.3 Hz), 1.80−1.25 (14H, m).

Step 6: Preparation of 7-Hydroxysulfamoylheptanoic Acid Phenylamide (6). Compound **6** was prepared from **61** obtained above using the procedure described for **12** (step 2) in 61% yield: mp 137−139 °C; ^{1}H NMR (DMSO-d_6, 400 MHz, δ; ppm) 9.85 (1H, broad s), 9.51 (1H, d, J = 3.2 Hz), 9.13 (1H, d, J = 3.2 Hz), 7.58 (2H, d, J = 8 Hz), 7.28 (2H, t, J = 7.8 Hz), 7.01 (1H, t, J = 7.3 Hz), 3.09 (2H, t, J = 7.6 Hz), 2.30 (2H, t, J = 7.3 Hz), 1.68 (2H, quintet, J = 8 Hz), 1.59 (2H, quintet, J = 7.6 Hz), 1.41 (2H, quintet, J = 7.8 Hz), 1.32 (2H, quintet, J = 7.1 Hz); Anal. ($C_{13}H_{20}N_2O_4S$·1/20H_2O) C, H, N.

Thioacetic acid S−(6-phenylcarbamoylhexyl) Ester (8a). Steps 1, 2, and 3: Preparation of 7-Bromoheptanoic Acid Phenylamide (64c). 7-Bromoheptanoic acid was prepared from **59** using the procedure described for **6** (step 4) in 99% yield. In this case, LiOH was used instead of NaOH: ^{1}H NMR ($CDCl_3$, 400 MHz, δ; ppm) 3.41 (2H, t, J = 6.8 Hz), 2.37 (2H, t, J = 7.3 Hz), 1.87 (2H, quintet, J = 6.8 Hz), 1.66 (2H, quintet, J = 7.6 Hz), 1.54−1.32 (4H, m).

To a suspension of 7-bromoheptanoic acid (2.64 g, 12.6 mmol) obtained above in CH_2Cl_2 (30 mL) were added oxalyl chloride (1.65 mL, 18.9 mmol) and a catalytic amount of DMF. The mixture was stirred at room temperature for 2 h. The solvent was removed by evaporation in vacuo to give acid chloride **62c**.

To a solution of aniline (3.50 g, 37.6 mmol) and triethylamine (5.30 mL, 38.1 mmol) in CH_2Cl_2 (40 mL) was added a solution of **62c** obtained above in CH_2Cl_2 (10 mL) dropwise cooling in an ice−water bath. The mixture was stirred at room temperature for 1 h. It was diluted with AcOEt and washed with aqueous saturated $NaHCO_3$, water, and brine, before being dried over $MgSO_4$. Filtration and concentration in vacuo and purification by silica gel flash chromatography (*n*-hexane/AcOEt = 3/1) gave 3.13 g (87%) of **64c**: ^{1}H NMR ($CDCl_3$, 400 MHz, δ; ppm) 7.51 (2H, d, J = 8.1 Hz), 7.32 (2H, t, J = 7.6 Hz), 7.15 (1H, broad s), 7.10 (1H, t, J = 7.6 Hz), 3.41 (2H, t, J = 6.8 Hz), 2.36 (2H, t, J = 7.3 Hz), 1.87 (2H, quintet, J = 7.1 Hz), 1.75 (2H, quintet, J = 7.3 Hz), 1.49 (2H, quintet, J = 7.6 Hz), 1.41 (2H, quintet, J = 6.8 Hz).

Step 4: Preparation of Thioacetic acid *S*-(6-Phenylcarbamoylhexyl) Ester (8a). Compound **8a** was prepared from **64c** obtained above using the procedure described for **15** in 98% yield: mp 80−81 °C; ^{1}H NMR ($CDCl_3$, 400 MHz, δ; ppm) 7.51 (2H, d, J = 8 Hz), 7.32 (2H, t, J = 7.3 Hz), 7.22

(1H, broad s), 7.10 (1H, t, J = 7.3 Hz), 2.86 (2H, t, J = 7.1 Hz), 2.35 (2H, t, J = 7.3 Hz), 2.32 (3H, s), 1.73 (2H, quintet, J = 7.1 Hz), 1.59 (2H, quintet, J = 7.1 Hz), 1.40 (4H, m); MS (EI) *m/z*: 279 (M^+); Anal. ($C_{15}H_{21}NO_2S$) C, H, N.

7-Mercaptoheptanoic Acid Phenylamide (7) and 7-(6-Phenylcarbamoylhexyldisulfanyl)heptanoic Acid Phenylamide (37). Compounds **7** and **37** were prepared from **8a** using the procedure described for **6** (step 4) in 87% and 4% yield, respectively.

7: mp 88−89 °C; ^{1}H NMR ($CDCl_3$, 400 MHz, δ; ppm) 7.51 (2H, d, J = 8 Hz), 7.32 (2H, t, J = 7.6 Hz), 7.12 (1H, broad s), 7.10 (1H, t, J = 7.1 Hz), 2.53 (2H, q, J = 7.3 Hz), 2.36 (2H, t, J = 7.6 Hz), 1.74 (2H, quintet, J = 7.1 Hz), 1.63 (2H, quintet, J = 7.1 Hz), 1.42 (4H, m), 1.33 (1H, t, J = 7.8 Hz); MS (EI) *m/z*: 237 (M^+); Anal. ($C_{13}H_{19}NOS$) C, H, N.

37: mp 105−107 °C; ^{1}H NMR ($CDCl_3$, 400 MHz, δ; ppm) 7.51 (4H, d, J = 8 Hz), 7.41 (2H, broad s), 7.30 (4H, t, J = 7.8 Hz), 7.09 (2H, t, J = 7.3 Hz), 2.68 (4H, t, J = 7.3 Hz), 2.36 (4H, t, J = 7.6 Hz), 1.74 (4H, quintet, J = 7.3 Hz), 1.69 (4H, quintet, J = 7.1 Hz), 1.50−1.34 (8H, m); MS (EI) *m/z*: 472 (M^+); Anal. ($C_{26}H_{36}N_2O_2S_2$) C, H, N.

Compounds**19**−**21**, **24**, **26**−**31**, and **32** were prepared from **62** and an appropriate aromatic amine using the procedure described for **8a** and **7**.

8-Mercaptooctanoic acid phenylamide (19): mp 84−86 °C; ^{1}H NMR ($CDCl_3$, 400 MHz, δ; ppm) 7.51 (2H, d, J = 8 Hz), 7.32 (2H, t, J = 7.6 Hz), 7.14 (1H, broad s), 7.10 (1H, t, J = 7.3 Hz), 2.52 (2H, q, J = 7.3 Hz), 2.35 (2H, t, J = 7.6 Hz), 1.73 (2H, quintet, J = 7.3 Hz), 1.61 (2H, quintet, J = 7.1 Hz), 1.46−1.34 (6H, m), 1.33 (1H, t, J = 7.8 Hz); MS (EI) *m/z*: 251 (M^+); Anal. ($C_{14}H_{21}NOS$) C, H, N.

6-Mercaptohexanoic acid phenylamide (20): mp 84−85 °C; ^{1}H NMR ($CDCl_3$, 400 MHz, δ; ppm) 7.51 (2H, d, J = 8.1 Hz), 7.32 (2H, t, J = 7.6 Hz), 7.16 (1H, broad s), 7.11 (1H, t, J = 7.8 Hz), 2.55 (2H, q, J = 7.1 Hz), 2.37 (2H, t, J = 7.3 Hz), 1.75 (2H, quintet, J = 7.8 Hz), 1.68 (2H, quintet, J = 7.6 Hz), 1.56−1.40 (2H, m), 1.35 (1H, t, J = 7.8 Hz); MS (EI) *m/z*: 223 (M^+); Anal. ($C_{12}H_{17}NOS$) C, H, N.

5-Mercaptopentanoic acid phenylamide (21): mp 120−121 °C; ^{1}H NMR ($CDCl_3$, 400 MHz, δ; ppm) 7.51 (2H, d, J = 7.6 Hz), 7.33 (2H, t, J = 8 Hz), 7.16 (1H, broad s), 7.11 (1H, t, J = 7.8 Hz), 2.58 (2H, q, J = 6.4 Hz), 2.39 (2H, t, J = 6.8 Hz), 1.85 (2H, quintet, J = 7.8 Hz), 1.71 (2H, quintet, J = 7.6 Hz), 1.39 (1H, t, J = 8 Hz); MS (EI) *m/z*: 209 (M^+); Anal. ($C_{11}H_{15}NOS\cdot 1/12H_2O$) C, H, N.

7-Mercaptoheptanoic acid (4-dimethylaminophenyl)amide (24): mp 121−122 °C; ^{1}H NMR ($CDCl_3$, 400 MHz, δ; ppm) 7.51 (2H, d, J = 9 Hz), 6.96 (1H, broad s), 6.70 (2H, d, J = 9 Hz), 2.91 (6H, s), 2.53 (2H, q, J = 7.3 Hz), 2.32 (2H, t, J = 7.3 Hz), 1.73 (2H, quintet, J = 7.4 Hz), 1.63 (2H, quintet, J = 7.6 Hz), 1.50−1.35 (4H, m), 1.33 (1H, t, J = 7.8 Hz); MS (EI) *m/z*: 280 (M^+); Anal. ($C_{15}H_{24}N_2OS$) C, H, N.

7-Mercaptoheptanoic acid 3-biphenylylamide (26): mp 91−92 °C; ^{1}H NMR ($CDCl_3$, 400 MHz, δ; ppm) 7.78 (1H, s), 7.59 (2H, d, J = 7.6 Hz), 7.49 (1H, d, J = 7.4 Hz), 7.47−7.30 (5H, m), 7.18 (1H, broad s), 2.53 (2H, q, J = 7.3 Hz), 2.39 (2H, t, J = 7.3 Hz), 1.76 (2H, quintet, J = 7.1 Hz), 1.64 (2H, quintet, J = 7.3 Hz), 1.50−1.37 (4H, m), 1.33 (1H, t, J = 7.6 Hz); MS (EI) *m/z*: 313 (M^+); Anal. ($C_{19}H_{23}NOS$) C, H, N.

7-Mercaptoheptanoic acid (4-phenoxyphenyl)amide (27): mp 87−89 °C; ^{1}H NMR ($CDCl_3$, 400 MHz, δ; ppm) 7.47 (2H, d, J = 8.8 Hz), 7.32 (2H, t, J = 7.8 Hz), 7.12 (1H, broad s), 7.08 (1H, t, J = 7.3 Hz), 6.98 (4H, d, J = 8.8 Hz), 2.53 (2H, q, J = 7.3 Hz), 2.36 (2H, t, J = 7.6 Hz), 1.75 (2H, quintet, J = 7.1 Hz), 1.64 (2H, quintet, J = 7.1 Hz), 1.50−1.37 (4H, m), 1.33 (1H, t, J = 7.8 Hz); MS (EI) *m/z*: 329 (M^+); Anal. ($C_{19}H_{23}NO_2S$) C, H, N.

7-Mercaptoheptanoic acid (3-phenoxyphenyl)amide (28): mp 68−69 °C; ^{1}H NMR ($CDCl_3$, 400 MHz, δ; ppm) 7.34 (2H, t, J = 7.6 Hz), 7.30−7.18 (3H, m), 7.16 (1H, broad s), 7.11 (1H, t, J = 7.2 Hz), 7.02 (2H, d, J = 8.5 Hz), 6.74 (1H, s), 2.52 (2H, q, J = 7.3 Hz), 2.33 (2H, t, J = 7.3 Hz), 1.71 (2H, quintet, J = 7.3 Hz), 1.62 (2H, quintet, J = 7.1 Hz), 1.50−1.34 (4H, m), 1.32 (1H, t, J = 7.6 Hz); MS (EI) *m/z*: 329 (M^+); Anal. ($C_{19}H_{23}NO_2S$) C, H, N.

7-Mercaptoheptanoic acid 3-pyridinylamide (29): mp 74−76 °C; ^{1}H NMR ($CDCl_3$, 400 MHz, δ; ppm) 8.54 (1H, d, J = 2.4 Hz), 8.35 (1H, d, J = 4.4 Hz), 8.19 (1H, d, J = 8.3 Hz), 7.31 (1H, broad s), 7.28 (1H, dd, J = 4.4, 8.3 Hz), 2.53 (2H, q, J = 7.1 Hz), 2.40 (2H, t, J = 7.3 Hz), 1.75 (2H, quintet, J = 7.6 Hz), 1.64 (2H, quintet, J = 7.1 Hz), 1.50−1.36 (4H, m), 1.33 (1H, t, J = 7.6 Hz); MS (EI) *m/z*: 237 (M^+); Anal. ($C_{12}H_{18}N_2OS$) C, H, N.

7-Mercaptoheptanoic acid 3-quinolinylamide (30): mp 75−76 °C; ^{1}H NMR ($CDCl_3$, 400 MHz, δ; ppm) 8.79 (1H, d, J = 2.7 Hz), 8.72 (1H, d, J = 2.7 Hz), 8.04 (1H, d, J = 8.3 Hz), 7.80 (1H, d, J = 8.3 Hz), 7.64 (1H, t, J = 7.1 Hz), 7.54 (1H, t, J = 7.1 Hz), 7.50 (1H, broad s), 2.54 (2H, q, J = 7.1 Hz), 2.47 (2H, t, J = 7.3 Hz), 1.80 (2H, quintet, J = 7.3 Hz), 1.64 (2H, quintet, J = 7.3 Hz), 1.53−1.37 (4H, m), 1.34 (1H, t, J = 7.8 Hz); MS (EI) *m/z*: 288 (M^+); Anal. ($C_{16}H_{20}N_2OS$) C, H, N.

7-Mercaptoheptanoic acid (4-phenyl-2-thiazolyl)amide (31): mp 149−150 °C; ^{1}H NMR ($CDCl_3$, 400 MHz, δ; ppm) 10.36 (1H, broad s), 7.83 (2H, d, J = 7.1 Hz), 7.43 (2H, t, J = 7.3 Hz), 7.16 (1H, s), 2.49 (2H, q, J = 7.1 Hz), 2.14 (2H, t, J = 7.6 Hz), 1.65−1.50 (4H, m), 1.32 (1H, t, J = 7.6 Hz), 1.30 (2H, quintet, J = 7.3 Hz), 1.15 (2H, quintet, J = 7.1 Hz); MS (EI) *m/z*: 320 (M^+); Anal. ($C_{16}H_{20}N_2OS_2\cdot 1/10H_2O$) C, H, N.

7-Mercaptoheptanoic acid 2-benzothiazolylamide (32): mp 141−142 °C; ^{1}H NMR ($CDCl_3$, 400 MHz, δ; ppm) 10.71 (1H, broad s), 7.86 (1H, d, J = 7.9 Hz), 7.77 (1H, d, J = 8 Hz), 7.46 (1H, t, J = 8.3 Hz), 7.34 (1H, t, J = 8.3 Hz), 2.49 (2H, t, J = 7.1 Hz), 2.48 (2H, q, J = 7.3 Hz), 1.72 (2H, quint, J = 7.6 Hz), 1.57 (2H, quint, J = 7.3 Hz), 1.40−1.25 (5H, m); MS (EI) *m/z*: 294 (M^+); Anal. ($C_{14}H_{18}N_2OS_2$) C, H, N.

7-Mercaptoheptanoic Acid 4-Biphenylylamide (25). Step 1: Preparation of 7-Bromoheptanoic Acid (4-Bromophenyl)amide (64a). Compound **64a** was prepared from **62c** and 4-bromoaniline using the procedure described for **8a** (step 3) in 86% yield: ^{1}H NMR ($CDCl_3$, 400 MHz, δ; ppm) 7.42 (4H, s), 7.14 (1H, broad s), 3.41 (2H, t, J = 6.6 Hz), 2.36 (2H, t, J = 7.6 Hz), 1.87 (2H, quintet, J = 7.1 Hz), 1.74 (2H, quintet, J = 7.3 Hz), 1.49 (2H, quintet, J = 7.3 Hz), 1.40 (2H, quintet, J = 6.8 Hz).

Step 2: Preparation of 7-Bromoheptanoic Acid 4-Biphenylylamide (64b). To a suspension of **64a** (500 mg, 1.38 mmol) obtained above in 1-methyl-2-pyrrolidinone (8 mL) and water (4 mL) were added phenylboronic acid (252 mg, 2.07 mmol), tetrakis(triphenylphosphine)palladium(0) (160 mg, 0.14 mmol), and $NaHCO_3$ (235 mg, 2.80 mmol). The mixture was heated at 80 °C for 1 h. The solution was diluted with AcOEt, washed with saturated aqueous $NaHCO_3$, water, and brine, and dried over Na_2SO_4. Filtration and concentration in vacuo and purification by silica gel flash chromatography (*n*-hexane/AcOEt = 3/1) gave 91 mg (18%) of **64b** as a white solid: ^{1}H NMR ($CDCl_3$, 400 MHz, δ; ppm) 7.65−7.50 (6H, m), 7.43 (2H, t, J = 7.6 Hz), 7.33 (1H, t, J = 7.1 Hz), 7.20 (1H, broad s), 3.42 (2H, t, J = 6.6 Hz), 2.39 (2H, t, J = 7.3 Hz), 1.88 (2H, quintet, J = 7.1 Hz), 1.77 (2H, quintet, J = 7.3 Hz), 1.50 (2H, quintet, J = 7.1 Hz), 1.43 (2H, quintet, J = 6.4 Hz).

Steps 3 and 4: Preparation of 7-Mercaptoheptanoic Acid 4-Biphenylylamide (25). Compound **25** was prepared from **64b** obtained above using the procedure described for **15** and **6** (step 4) in 48% yield: mp 114−115 °C; ^{1}H NMR ($CDCl_3$, 400 MHz, δ; ppm) 7.64−7.52 (6H, m), 7.43 (2H, t, J = 7.6 Hz), 7.33 (1H, t, J = 7.3 Hz), 7.17 (1H, broad s), 2.54 (2H, q, J = 7.4 Hz), 2.39 (2H, t, J = 7.3 Hz), 1.76 (2H, quintet, J = 7.3 Hz), 1.64 (2H, quintet, J = 7.3 Hz), 1.52−1.37 (4H, m), 1.34 (1H, t, J = 7.6 Hz); MS (EI) *m/z*: 313 (M^+); Anal. ($C_{19}H_{23}NOS\cdot 1/5H_2O$) C, H, N.

7-Methylsulfanylheptanoic Acid Phenylamide (9). To a solution of **64c** (300 mg, 1.06 mmol) in EtOH (10 mL) was added methylmercaptan sodium salt (15% in water, 1.50 g, 3.21 mmol), and the solution was stirred at room temperature for 5 h. The reaction mixture was diluted with AcOEt, washed with water and brine, and dried over $MgSO_4$. Filtration and concentration in vacuo and purification by silica gel flash

chromatography (*n*-hexane/AcOEt = 2/1) gave 262 mg (99%) of **9** as a crude solid. The solid was recrystallized from *n*-hexane−AcOEt and collected by filtration to give 217 mg of **9** as a colorless crystal: mp 50−51 °C; ^{1}H NMR ($CDCl_3$, 400 MHz, δ; ppm) 7.51 (2H, d, J = 8 Hz), 7.32 (2H, t, J = 7.8 Hz), 7.16 (1H, broad s), 7.10 (1H, t, J = 7.6 Hz), 2.49 (2H, t, J = 7.1 Hz), 2.36 (2H, t, J = 7.3 Hz), 2.09 (3H, s), 1.74 (2H, quintet, J = 7.3 Hz), 1.61 (2H, quintet, J = 7.3 Hz), 1.42 (4H, m); MS (EI) *m/z*: 251 (M^+); Anal. ($C_{14}H_{21}NOS$) C, H, N.

7-Methanesulfonylheptanoic Acid Phenylamide (11). To a solution of **9** (80 mg, 0.32 mmol) in CH_2Cl_2 (3 mL) was added 3-chloroperoxybenzoic acid (65%, 180 mg, 0.68 mmol). The mixture was stirred overnight at room temperature. Next, saturated aqueous $NaHCO_3$ and saturated aqueous $Na_2S_2O_3$ were added, and the mixture was stirred at room temperature for 1 h. It was then poured into water and extracted with $CHCl_3$. The $CHCl_3$ layer was separated, washed with water and brine, and dried over Na_2SO_4. Filtration and concentration in vacuo and separation by silica gel flash chromatography (*n*-hexane/AcOEt = 1/3) gave 63 mg (70%) of **11** as a crude solid. The solid was recrystallized from *n*-hexane-AcOEt and collected by filtration to give 50 mg of **11** as a colorless crystal: mp 121−123 °C; ^{1}H NMR ($CDCl_3$, 400 MHz, δ; ppm) 7.51 (2H, d, J = 7.8 Hz), 7.32 (2H, t, J = 7.6 Hz), 7.17 (1H, brs), 7.11 (1H, t, J = 7.3 Hz), 3.01 (2H, t, J = 7.8 Hz), 2.89 (3H, s), 2.37 (2H, t, J = 7.3 Hz), 1.88 (2H, quint, J = 7.6 Hz), 1.76 (2H, quint, J = 7.6 Hz), 1.60−1.35 (4H, m); MS (EI) *m/z*: 283 (M^+); Anal. ($C_{14}H_{21}NO_3S$) C, H, N.

6-Phenoxy-1-hexanethiol (22). Step 1: Preparation of 6-Phenoxy-1-hexanol (67). To a solution of phenol (2.10 g, 22.31 mmol) and 6-bromo-1-hexanol (**65**, 2.00 g, 11.05 mmol) in DMF (30 mL) was added K_2CO_3 (3.10 g, 22.4 mmol), and the mixture was stirred at 80 °C for 1 h. The reaction mixture was diluted with AcOEt and washed with water and brine, before being dried over Na_2SO_4. Filtration and concentration in vacuo and purification by silica gel flash chromatography (*n*-hexane/AcOEt = 2/1) gave 2.06 g (96%) of **67** as a white solid: ^{1}H NMR ($CDCl_3$, 400 MHz, δ; ppm) 7.28 (2H, t, J = 7.8 Hz), 6.93 (1H, t, J = 7.3 Hz), 6.89 (2H, d, J = 8.6 Hz), 3.96 (2H, t, J = 6.6 Hz), 3.67 (2H, m), 1.80 (2H, quintet, J = 6.8 Hz), 1.61 (2H, quintet, J = 7.3 Hz), 1.56−1.36 (4H, m), 1.27 (1H, m).

Step 2: Preparation of (6-Bromohexyloxy)benzene. To a solution of **67** (1.75 g, 9.01 mmol) obtained above and carbon tetrabromide (3.00 g, 9.05 mmol) in CH_2Cl_2 (50 mL) was added triphenylphosphine (2.60 g, 9.91 mmol) with cooling in an ice−water bath. The solution was stirred at room temperature for 1 h and concentrated in vacuo. To the residue was added *n*-hexane (30 mL), and the slurry was filtered. After the solid was washed with *n*-hexane (10 mL), the combined filtrates were concentrated in vacuo. The residue was purified by silica gel flash chromatography (*n*-hexane/AcOEt = 1/30) to give 1.45 g (63%) of (6-bromohexyloxy)benzene as a colorless oil: ^{1}H NMR ($CDCl_3$, 400 MHz, δ; ppm) 7.28 (2H, t, J = 7.6 Hz), 6.93 (1H, t, J = 7.3 Hz), 6.89 (2H, d, J = 8.5 Hz), 3.96 (2H, t, J = 6.3 Hz), 3.43 (2H, t, J = 6.8 Hz), 1.90 (2H, quintet, J = 6.8 Hz), 1.80 (2H, quintet, J = 6.4 Hz), 1.56−1.46 (4H, m).

Steps 3 and 4: Preparation of 6-Phenoxy-1-hexanethiol (22). Compound **22** was prepared from (6-bromohexyloxy)-benzene obtained above using the procedure described for **15** and **6** (step 4) in 45% yield: colorless oil; ^{1}H NMR ($CDCl_3$, 400 MHz, δ; ppm) 7.28 (2H, t, J = 7.3 Hz), 6.93 (1H, t, J = 7.6 Hz), 6.89 (2H, d, J = 7.8 Hz), 3.96 (2H, t, J = 6.4 Hz), 2.54 (2H, q, J = 7.1 Hz), 1.79 (2H, quintet, J = 6.6 Hz), 1.65 (2H, quintet, J = 6.8 Hz), 1.54−1.44 (4H, m), 1.34 (1H, t, J = 7.8 Hz); MS (EI) *m/z*: 210 (M^+); HRMS calcd for $C_{12}H_{18}OS$ 210.108, found 210.108.

Compounds**23**, **33**−**35**, and **36** were prepared from an appropriate aromatic carboxylic acid and 6-amino-1-hexanol (**66**) using the procedure described for **13**, **22** (step 2), **15**, and **6** (step 4).

***N*-(6-Mercaptohexyl)benzamide (23)**: mp 43−44 °C; ^{1}H NMR ($CDCl_3$, 400 MHz, δ; ppm) 7.77 (2H, d, J = 7.2 Hz), 7.50 (1H, t, J = 7.2 Hz), 7.43 (2H, t, J = 6.8 Hz), 6.20 (1H, broad s), 3.47 (2H, q, J = 6.4 Hz), 2.54 (2H, q, J = 7.6 Hz), 1.68−1.58 (4H, m), 1.50−1.36 (4H, m), 1.52−1.37 (4H, m), 1.34 (1H, t, J = 7.8 Hz); MS (EI) *m/z*: 237 (M^+); Anal. ($C_{13}H_{19}NOS\cdot 1/6H_2O$) C, H, N.

4-Dimethylamino-*N*-(6-mercaptohexyl)benzamide (33): mp 103−104 °C; ^{1}H NMR ($CDCl_3$, 400 MHz, δ; ppm) 7.66 (2H, t, J = 8.8 Hz), 6.67 (2H, d, J = 8.8 Hz), 5.95 (1H, s), 3.43 (2H, q, J = 6.4 Hz), 3.02 (1H, s), 2.53 (2H, q, J = 7.2 Hz), 1.67−1.54 (4H, m), 1.49−1.36 (4H, m), 1.32 (1H, t, J = 8 Hz); MS (EI) *m/z*: 280 (M^+); Anal. ($C_{15}H_{24}N_2OS$) C, H, N.

Naphthalene-2-carboxylic acid (6-mercaptohexyl)amide (34): mp 76−78 °C; ^{1}H NMR ($CDCl_3$, 400 MHz, δ; ppm) 8.28 (1H, s), 7.94−7.85 (3H, m), 7.82 (1H, d, J = 6.8 Hz), 7.58−7.53 (2H, m), 6.27 (1H, s), 3.52 (2H, q, J = 6.8 Hz), 2.54 (2H, q, J = 7.6 Hz), 1.70−1.62 (4H, m), 1.52−1.36 (4H, m), 1.34 (1H, t, J = 7.8 Hz); MS (EI) *m/z*: 287 (M^+); Anal. ($C_{17}H_{21}NOS$) C, H, N.

Benzofuran-2-carboxylic acid (6-mercaptohexyl)amide (35): mp 72−73 °C; ^{1}H NMR ($CDCl_3$, 400 MHz, δ; ppm) 7.68 (1H, d, J = 8 Hz), 7.50 (1H, d, J = 8.4 Hz), 7.46 (1H, s), 7.41 (1H, t, J = 8.4 Hz), 7.30 (1H, t, J = 8 Hz), 6.64 (1H, s), 3.49 (2H, q, J = 7.2 Hz), 2.54 (2H, q, J = 7.2 Hz), 1.72−1.58 (4H, m), 1.52−1.38 (4H, m), 1.34 (1H, t, J = 7.8 Hz); MS (EI) *m/z*: 277 (M^+); Anal. ($C_{15}H_{19}NO_2S$) C, H, N.

Indole-2-carboxylic acid (6-mercaptohexyl)amide (36): mp 128−130 °C; ^{1}H NMR ($CDCl_3$, 400 MHz, δ; ppm) 9.12 (1H, broad s), 7.65 (1H, d, J = 8 Hz), 7.44 (1H, d, J = 8.4 Hz), 7.29 (1H, t, J = 8 Hz), 7.14 (1H, t, J = 6.6 Hz), 6.82 (1H, s), 6.13 (1H, broad s), 3.49 (2H, q, J = 6.8 Hz), 2.54 (2H, q, J = 7.6 Hz), 1.70−1.60 (4H, m), 1.45−1.40 (4H, m), 1.34 (1H, t, J = 7.8 Hz); MS (EI) *m/z*: 276 (M^+); Anal. ($C_{15}H_{20}N_2OS$) C, H, N.

Thiopropionic Acid *S*-(6-Phenylcarbamoylhexyl) Ester (38). To a solution of **7** (200 mg, 0.84 mmol) and a catalytic amount of 4-(dimethylamino)pyridine in CH_2Cl_2 (2 mL) and pyridine (0.5 mL) was added propionyl chloride (220 μL, 2.53 mmol). The mixture was stirred at room temperature for 30 min and then diluted with AcOEt. The solution was washed with water and brine and dried over Na_2SO_4. Filtration and concentration in vacuo and separation by silica gel flash chromatography (*n*-hexane/AcOEt = 3/1) gave 238 mg (96%) of **38** as a crude solid. The solid was recrystallized from *n*-hexane-AcOEt and collected by filtration to give 184 mg of **38** as a colorless crystal: mp 54−55 °C; ^{1}H NMR ($CDCl_3$, 500 MHz, δ; ppm) 7.52 (2H, d, J = 7.9 Hz), 7.32 (2H, t, J = 7.9 Hz), 7.21 (1H, broad s), 7.10 (1H, t, J = 7.3 Hz), 2.86 (2H, t, J = 7.4 Hz), 2.57 (2H, q, J = 7.7 Hz), 2.35 (2H, t, J = 7.6 Hz), 1.74 (2H, quintet, J = 7.3 Hz), 1.59 (2H, quintet, J = 7.3 Hz), 1.46−1.33 (4H, m), 1.18 (3H, t, J = 7.7 Hz); MS (EI) *m/z*: 293 (M^+); Anal. ($C_{16}H_{23}NO_2S$) C, H, N.

Compounds **39**−**45**, **47**−**54**, and **55** were prepared from the corresponding thiols and an appropriate acid chloride using the procedure described for **38**.

Thiobutyric acid *S*-(6-phenylcarbamoylhexyl) ester (39): mp 45−46 °C; ^{1}H NMR ($CDCl_3$, 500 MHz, δ; ppm) 7.52 (2H, d, J = 8 Hz), 7.32 (2H, t, J = 7.6 Hz), 7.21 (1H, broad s), 7.10 (1H, t, J = 7.3 Hz), 2.86 (2H, t, J = 7 Hz), 2.52 (2H, t, J = 7.3 Hz), 2.35 (2H, t, J = 7.4 Hz), 1.73 (2H, quintet, J = 7.4 Hz), 1.69 (2H, sextet, J = 7.7 Hz), 1.59 (2H, quintet, J = 7.4 Hz), 1.48−1.33 (4H, m), 0.95 (3H, t, J = 7.3 Hz); MS (EI) *m/z*: 307 (M^+); Anal. ($C_{17}H_{25}NO_2S$) C, H, N.

Thioisobutyric acid *S*-(6-phenylcarbamoylhexyl) ester (40): mp 44−45 °C; ^{1}H NMR ($CDCl_3$, 400 MHz, δ; ppm) 7.52 (2H, d, J = 8 Hz), 7.32 (2H, t, J = 7.8 Hz), 7.22 (1H, broad s), 7.10 (1H, t, J = 7.3 Hz), 2.85 (2H, t, J = 7.3 Hz), 2.73 (1H, septet, J = 7 Hz), 2.35 (2H, t, J = 7.3 Hz), 1.73 (2H, quintet, J = 7.3 Hz), 1.59 (2H, quintet, J = 7.3 Hz), 1.46−1.36 (4H, m), 1.19 (6H, d, J = 7.6 Hz); MS (EI) *m/z*: 307 (M^+); Anal. ($C_{17}H_{25}NO_2S$) C, H, N.

2,2-Dimethylthiopropionic acid *S*-(6-phenylcarbamoylhexyl) ester (41): mp 57−59 °C; ^{1}H NMR ($CDCl_3$, 400 MHz, δ; ppm) 7.52 (2H, d, J = 8.1 Hz), 7.32 (2H, t, J = 7.6 Hz), 7.20 (1H, broad s), 7.10 (1H, t, J = 7.6 Hz), 2.82 (2H, t, J = 7.3 Hz), 2.35 (2H, t, J = 7.3 Hz), 1.73 (2H, quintet, J = 7.3

Hz), 1.58 (2H, quintet, J = 7.3 Hz), 1.46−1.36 (4H, m), 1.23 (9H, s); MS (EI) *m/z*: 321 (M^+); Anal. ($C_{18}H_{27}NO_2S$) C, H, N.

Cyclopropanecarbothioic acid *S*-(6-phenylcarbamoylhexyl) ester (42): mp 64−65 °C; ^{1}H NMR ($CDCl_3$, 500 MHz, δ; ppm) 7.52 (2H, d, J = 8.3 Hz), 7.32 (2H, t, J = 7.6 Hz), 7.22 (1H, broad s), 7.10 (1H, t, J = 7.3 Hz), 2.89 (2H, t, J = 7.3 Hz), 2.35 (2H, t, J = 7.3 Hz), 2.01 (1H, m), 1.73 (2H, quintet, J = 7 Hz), 1.59 (2H, quintet, J = 7.3 Hz), 1.45−1.35 (4H, m), 1.15 (2H, m), 0.94 (2H, m); MS (EI) *m/z*: 305 (M^+); Anal. ($C_{17}H_{23}NO_2S$) C, H, N.

Cyclopentanecarbothioic acid *S*-(6-phenylcarbamoylhexyl) ester (43): mp 59−60 °C; ^{1}H NMR ($CDCl_3$, 500 MHz, δ; ppm) 7.52 (2H, d, J = 7.9 Hz), 7.32 (2H, t, J = 7.9 Hz), 7.21 (1H, broad s), 7.10 (1H, t, J = 7.3 Hz), 2.97 (1H, quintet, J = 8 Hz), 2.85 (2H, t, J = 7.4 Hz), 2.35 (2H, t, J = 7.7 Hz), 1.93−1.67 (8H, m), 1.63−1.52 (4H, m), 1.47−1.33 (4H, m); MS (EI) *m/z*: 333 (M^+); Anal. ($C_{19}H_{27}NO_2S$) C, H, N.

Thiobenzoic acid *S*-(6-phenylcarbamoylhexyl) ester (44): mp 107−109 °C; ^{1}H NMR ($CDCl_3$, 400 MHz, δ; ppm) 7.97 (2H, d, J = 7.3 Hz), 7.57 (1H, t, J = 7.3 Hz), 7.52 (2H, d, J = 7.8 Hz), 7.45 (2H, t, J = 7.8 Hz), 7.31 (2H, t, J = 7.6 Hz), 7.21 (1H, broad s), 7.10 (1H, t, J = 7.3 Hz), 3.07 (2H, t, J = 7.3 Hz), 2.36 (2H, t, J = 7.3 Hz), 1.75 (2H, quintet, J = 7.3 Hz), 1.70 (2H, quintet, J = 7.3 Hz), 1.54−1.36 (4H, m); MS (EI) *m/z*: 341 (M^+); Anal. ($C_{20}H_{23}NO_2S$) C, H, N.

4-Nitrothiobenzoic acid *S*-(6-phenylcarbamoylhexyl) ester (45): mp 117−118 °C; ^{1}H NMR ($CDCl_3$, 400 MHz, δ; ppm) 8.30 (2H, d, J = 8.8 Hz), 8.11 (2H, d, J = 8.6 Hz), 7.51 (2H, d, J = 8.1 Hz), 7.32 (2H, t, J = 7.8 Hz), 7.16 (1H, broad s), 7.10 (1H, t, J = 7.3 Hz), 3.12 (2H, t, J = 7.3 Hz), 2.37 (2H, t, J = 7.3 Hz), 1.76 (2H, quintet, J = 7.6 Hz), 1.72 (2H, quintet, J = 7.3 Hz), 1.54−1.38 (4H, m); MS (EI) *m/z*: 386 (M^+); Anal. ($C_{20}H_{22}N_2O_4S$) C, H, N.

Thioisobutyric acid *S*-[6-(3-biphenylylcarbamoyl)-hexyl] ester (47): mp 73−74 °C; ^{1}H NMR ($CDCl_3$, 500 MHz, δ; ppm) 7.79 (1H, s), 7.59 (2H, d, J = 7.4 Hz), 7.50 (1H, d, J = 8.3 Hz), 7.43 (2H, t, J = 7.3 Hz), 7.39 (1H, t, J = 8 Hz), 7.35 (1H, t, J = 7.3 Hz), 7.34 (1H, d, J = 7.3 Hz), 7.28 (1H, broad s), 2.85 (2H, t, J = 7.3 Hz), 2.73 (1H, septet, J = 6.8 Hz), 2.38 (2H, t, J = 7.3 Hz), 1.75 (2H, quintet, J = 7.6 Hz), 1.58 (2H, quintet, J = 7.3 Hz), 1.49−1.35 (4H, m), 1.18 (6H, d, J = 7.1 Hz); MS (EI) *m/z*: 390 (M^+); Anal. ($C_{23}H_{29}NO_2S$) C, H, N.

Thioisobutyric acid *S*-[6-(3-phenoxyphenylcarbamoyl)hexyl] ester (48): colorless oil; ^{1}H NMR ($CDCl_3$, 500 MHz, δ; ppm) 7.34 (2H, t, J = 7.6 Hz), 7.30−7.15 (4H, m), 7.11 (1H, t, J = 7.4 Hz), 7.02 (2H, d, J = 7.6 Hz), 6.74 (1H, d, J = 7.3 Hz), 2.84 (2H, t, J = 7.3 Hz), 2.73 (1H, septet, J = 7 Hz), 2.32 (2H, t, J = 7.3 Hz), 1.71 (2H, quintet, J = 7.4 Hz), 1.57 (2H, quintet, J = 7.4 Hz), 1.45−1.33 (4H, m), 1.18 (6H, d, J = 7 Hz); MS (EI) *m/z*: 399 (M^+); HRMS calcd for $C_{23}H_{29}NO_3S$ 399.187, found 399.191.

Thioisobutyric acid *S*-[6-(3-pyridinylcarbamoyl)hexyl] ester (49): mp 47−48 °C; ^{1}H NMR ($CDCl_3$, 500 MHz, δ; ppm) 8.55 (1H, d, J = 2.8 Hz), 8.34 (1H, d, J = 4.6 Hz), 8.21 (1H, d, J = 8.5 Hz), 7.56 (1H, broad s), 7.28 (1H, dd, J = 4.6, 8.3 Hz), 2.85 (2H, t, J = 7 Hz), 2.74 (1H, septet, J = 7 Hz), 2.39 (2H, t, J = 7.6 Hz), 1.75 (2H, quintet, J = 7.4 Hz), 1.59 (2H, quintet, J = 7.1 Hz), 1.45−1.35 (4H, m), 1.19 (6H, d, J = 6.8 Hz); MS (EI) *m/z*: 308 (M^+); Anal. ($C_{16}H_{24}N_2O_2S$) C, H, N.

Thioisobutyric acid *S*-[6-(3-quinolinylcarbamoyl)-hexyl] ester (50): mp 67−68 °C; ^{1}H NMR ($CDCl_3$, 500 MHz, δ; ppm) 8.81 (1H, s), 8.73 (1H, d, J = 2.8 Hz), 8.03 (1H, d, J = 8.6 Hz), 7.80 (1H, d, J = 8.2 Hz), 7.70 (1H, broad s), 7.63 (1H, t, J = 7.1 Hz), 7.54 (1H, t, J = 7.3 Hz), 2.86 (2H, t, J = 7.3 Hz), 2.74 (1H, septet, J = 7 Hz), 2.46 (2H, t, J = 7.6 Hz), 1.79 (2H, quintet, J = 7.3 Hz), 1.60 (2H, quintet, J = 7.3 Hz), 1.50−1.35 (4H, m), 1.19 (6H, d, J = 6.7 Hz); MS (EI) *m/z*: 358 (M^+); Anal. ($C_{20}H_{26}N_2O_2S$) C, H, N.

Thioisobutyric acid *S*-[6-(4-phenyl-2-thiazolylcarbamoyl)hexyl] ester (51): mp 127−128 °C; ^{1}H NMR ($CDCl_3$, 500 MHz, δ; ppm) 10.48 (1H, broad s), 7.83 (2H, d, J = 7.3 Hz), 7.43 (2H, t, J = 7.3 Hz), 7.34 (1H, t, J = 7.4 Hz), 7.16 (1H, s), 2.81 (2H, t, J = 7.3 Hz), 2.74 (1H, septet, J = 7 Hz), 2.11 (2H, t, J = 7.6 Hz), 1.56 (2H, quintet, J = 7.6 Hz), 1.50 (2H, quintet, J = 7.3 Hz), 1.25 (2H, quintet, J = 7.6 Hz), 1.19 (6H, d, J = 7 Hz), 1.13 (2H, quintet, J = 7.3 Hz); MS (EI) *m/z*: 383 (M^+); Anal. ($C_{20}H_{26}N_2O_2S_2$) C, H, N.

Thioisobutyric acid *S*-[6-(2-benzothiazolylcarbamoyl)-hexyl] ester (52): mp 106−107 °C; ^{1}H NMR ($CDCl_3$, 500 MHz, δ; ppm) 10.41 (1H, broad s), 7.85 (1H, d, J = 7.4 Hz), 7.77 (1H, d, J = 7.9 Hz), 7.46 (1H, dt, J = 1.2, 7.1 Hz), 7.34 (1H, dt, J = 1, 7.3 Hz), 2.81 (2H, t, J = 7.4 Hz), 2.73 (1H, septet, J = 7.1 Hz), 2.47 (2H, t, J = 7.7 Hz), 1.72 (2H, quintet, J = 7.3 Hz), 1.53 (2H, quintet, J = 7.1 Hz), 1.38−1.27 (4H, m), 1.18 (6H, d, J = 7 Hz); MS (EI) *m/z*: 364 (M^+); Anal. ($C_{18}H_{24}N_2O_2S_2$) C, H, N.

Thioisobutyric acid *S*-{6-[(2-naphthalenecarbonyl)-amino]hexyl} ester (53): mp 70−71 °C; ^{1}H NMR ($CDCl_3$, 500 MHz, δ; ppm) 8.29 (1H, s), 7.93 (1H, d, J = 7.1 Hz), 7.90 (1H, d, J = 7.3 Hz), 7.88 (1H, d, J = 7.3 Hz), 7.84 (1H, d, J = 7 Hz), 7.57 (1H, t, J = 6.7 Hz), 7.54 (1H, t, J = 6.7 Hz), 6.36 (1H, broad s), 3.51 (2H, q, J = 6.4 Hz), 2.87 (2H, t, J = 7.3 Hz), 2.73 (1H, septet, J = 6.7 Hz), 1.67 (2H, quintet, J = 7.1 Hz), 1.60 (2H, quintet, J = 6.7 Hz), 1.50−1.38 (4H, m), 1.18 (6H, d, J = 6.8 Hz); MS (EI) *m/z*: 357 (M^+); Anal. ($C_{21}H_{27}NO_2S$) C, H, N.

Thioisobutyric acid *S*-{6-[(2-benzofurancarbonyl)amino]hexyl} ester (54): mp 67−68 °C; ^{1}H NMR ($CDCl_3$, 500 MHz, δ; ppm) 7.67 (1H, d, J = 7.7 Hz), 7.50 (1H, d, J = 7.6 Hz), 7.46 (1H, d, J = 1 Hz), 7.41 (1H, dt, J = 1.2, 7.3 Hz), 7.29 (1H, t, J = 7.6 Hz), 6.66 (1H, broad s), 3.48 (2H, q, J = 7 Hz), 2.86 (2H, t, J = 7.4 Hz), 2.73 (1H, septet, J = 7.1 Hz), 1.66 (2H, quintet, J = 7 Hz), 1.59 (2H, quintet, J = 7 Hz), 1.48−1.37 (4H, m), 1.18 (6H, d, J = 6.7 Hz); MS (EI) *m/z*: 347 (M^+); Anal. ($C_{19}H_{25}NO_3S$) C, H, N.

Thioisobutyric acid *S*-{6-[(1*H*-2-indolecarbonyl)amino]hexyl} ester (55): mp 142−143 °C; ^{1}H NMR ($CDCl_3$, 500 MHz, δ; ppm) 9.37 (1H, broad s), 7.65 (1H, d, J = 7.3 Hz), 7.44 (1H, d, J = 7.6 Hz), 7.29 (1H, t, J = 7 Hz), 7.14 (1H, t, J = 7.9 Hz), 6.86 (1H, s), 6.30 (1H, broad s), 3.49 (2H, q, J = 6.1 Hz), 2.87 (2H, t, J = 7.1 Hz), 2.74 (1H, septet, J = 7 Hz), 1.65 (2H, quintet, J = 7 Hz), 1.60 (2H, quintet, J = 7 Hz), 1.50−1.36 (4H, m), 1.19 (6H, d, J = 7 Hz); MS (EI) *m/z*: 346 (M^+); Anal. ($C_{19}H_{26}N_2O_2S$) C, H, N.

2,2-Dimethylpropionic Acid 6-Phenylcarbamoylhexylsulfanylmethyl Ester (46). To a suspension of sodium hydride (60%, 40.0 mg, 1.00 mmol) in DMF (2 mL) was added a solution of **7** (200 mg, 0.84 mmol) in DMF (3 mL) dropwise with cooling in an ice−water bath. The mixture was stirred for 30 min at 0 °C, and a solution of chloromethyl pivalate (134 μL, 0.93 mmol) in DMF (2 mL) was added at 0 °C. The solution was stirred at room temperature for 1 h. The reaction mixture was poured into ice−water and extracted with AcOEt. The AcOEt layer was separated, washed with water and brine, and dried over Na_2SO_4. Filtration and concentration in vacuo and purification by silica gel flash chromatography (*n*-hexane/AcOEt = 4/1) gave 93 mg (32%) of **46** as a colorless oil: ^{1}H NMR ($CDCl_3$, 400 MHz, δ; ppm) 7.51 (2H, d, J = 7.8 Hz), 7.32 (2H, t, J = 7.6 Hz), 7.16 (1H, broad s), 7.10 (1H, t, J = 7.3 Hz), 5.41 (2H, s), 2.65 (2H, t, J = 7.3 Hz), 2.36 (2H, t, J = 7.6 Hz), 1.74 (2H, quintet, J = 7.1 Hz), 1.66 (2H, quintet, J = 7.1 Hz), 1.50−1.36 (4H, m), 1.21 (9H, s); MS (EI) *m/z*: 351 (M^+); HRMS calcd for $C_{19}H_{29}NO_3S$ 351.187, found 351.189.

Biology. Enzyme Assays. The assay of HDAC activity was performed using an HDAC fluorescent activity assay/drug discovery kit (AK-500, BIOMOL Research Laboratories). HeLa nuclear extracts (0.5 μL/well) were incubated at 37 °C with 25 μM of Fluor de Lys substrate and various concentrations of samples. Reactions were stopped after 30 min by adding Fluor de Lys Developer with trichostatin A which stops further deacetylation. Then, 15 min after addition of this developer, the fluorescence of the wells was measured on a fluorometric reader with excitation set at 360 nm and emission detection set at 460 nm, and the % inhibition was calculated from the fluorescence readings of inhibited wells relative to those of control wells. The concentration of compound which results in 50% inhibition was determined by plotting the log[Inh] versus the logit function of the % inhibition. IC_{50} values are

determined using a regression analysis of the concentration/inhibition data.

Lineweaver−Burk Double-Reciprocal Plot Analysis. The assay of HDAC activity was performed using an HDAC fluorescent activity assay/drug discovery kit (AK-500, BIOMOL Research Laboratories). HeLa nuclear extracts (0.5 μL/well) were incubated at 37 °C with Fluor de Lys substrate (50, 100, 200, or 400 μM) in the presence of 0, 0.03, 0.1, or 0.3 μM of compound **7**. Reactions were stopped after 10 min by adding Fluor de Lys Developer with trichostatin A which stops further deacetylation. Then, 15 min after addition of this developer, the fluorescence of the wells was measured on a fluorometric reader with excitation set at 360 nm and emission detection set at 460 nm.

Monolayer Growth Inhibition Assay. Cancer cells were plated in 96-well plates at initial densities of 1500 cells/well and incubated at 37 °C. After 24 h, cells were exposed to test compounds at various concentrations in 10% FBS-supplemented RPMI-1640 medium at 37 °C in 5% CO_2 for 48 h. The medium was removed and replaced with 200 μL of 0.5 mg/mL of Methylene Blue in RPMI-1640 medium, and cells were incubated at room temperature for 30 min. Supernatants were removed from the wells, and Methylene Blue dye was dissolved in 100 μL/well of 3% aqueous HCl. Absorbance was determined on a microplate reader (BioRad) at 660 nm.

Western Blot Analysis. HCT-116 cells (purchased from ATCC) (1×10^6) treated for 8 h with SAHA and compound **51** at the indicated concentrations in 10% FBS-supplemented McCoy's 5A medium were collected and sonicated. Protein concentrations of the lysates were determined by using a Bradford protein assay kit (Bio-Rad Laboratories); equivalent amounts of proteins from each lysate were resolved in 15% SDS-polyacrylamide gel and then transferred onto nitrocellulose membranes (Bio-Rad Laboratories). After blocking with Tris-buffered saline (TBS) containing 0.1% Tween 20 (TBST) containing 3% skim milk for 30 min, the transblotted membrane was incubated with hyperacetylated histone H4 antibody (Upstate Biotechnology) (1:2000) or $p21^{WAF1/CIP1}$ antibody (Medical and Biological Laboratories) (1: 200) in TBST containing 3% skim milk at 4 °C overnight. After treatment with the primary antibody, the membrane was washed twice with water for anti- hyperacetylated histone H4, or three times with TBS for anti-$p21^{WAF1/CIP1}$, then incubated with goat anti-rabbit or anti-mouse IgG-horseradish peroxidase conjugates (1:10000 or 1:5000) for 1.5 h at room temperature and washed twice with water for anti- hyperacetylated histone H4, or three times with TBS for anti-$p21^{WAF1/CIP1}$. The immunoblots were visualized by enhanced chemiluminescence.

Molecular Modeling. Docking and subsequent scoring were performed using Macromodel 8.1 software. Coordinates of HDAC8 complexed with MS344 were taken from the Brookhaven Protein Data Bank (PDB code 1T67) and hydrogen atoms were added computationally at appropriate positions. The structures of SAHA and compound **7** bound to HDAC8 were constructed by molecular mechanics (MM) energy minimization. The starting positions of SAHA and compound **7** were determined manually: the benzene ring and the linker parts were superimposed in the active site onto its crystallographic MS344 counterpart. The conformations of SAHA and compound **7** in the active site were minimized by a MM calculation based upon the OPLS-AA force field with each parameter set as follows: solvent: water, method: LBFGS, max. no. iterations: 10 000, converge on: gradient, convergence threshold: 0.05.

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Supporting Information Available: Results of the elemental analysis of **4**−**21**, **23**−**45**, **47**, **49**−**54**, and **55** are reported. This material is available free of charge via the Internet at http://pubs.acs.org.

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